

Nano-assemblies on materials :

*Mechanically responding
functionalized polyelectrolyte multilayer films*

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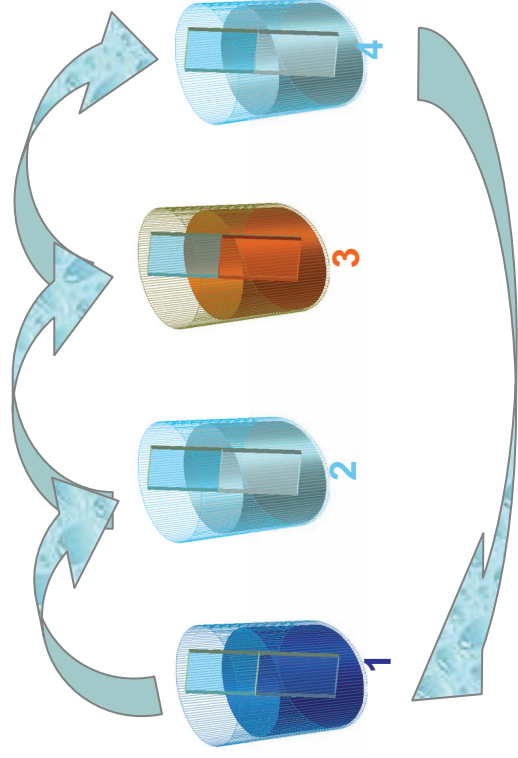
One of the main objective :

Modify biomaterial surfaces to provide them

- ➔ *Biomimetic properties*
 - ➔ *Multifunctional properties*
 - ➔ *Sequential release of drugs*
 - ➔ *Cover all materials*
- + Simple method*
 - + Structures well defined*

*Approach = polyelectrolyte multilayer film coatings
Layer-by-Layer method (LbL)*

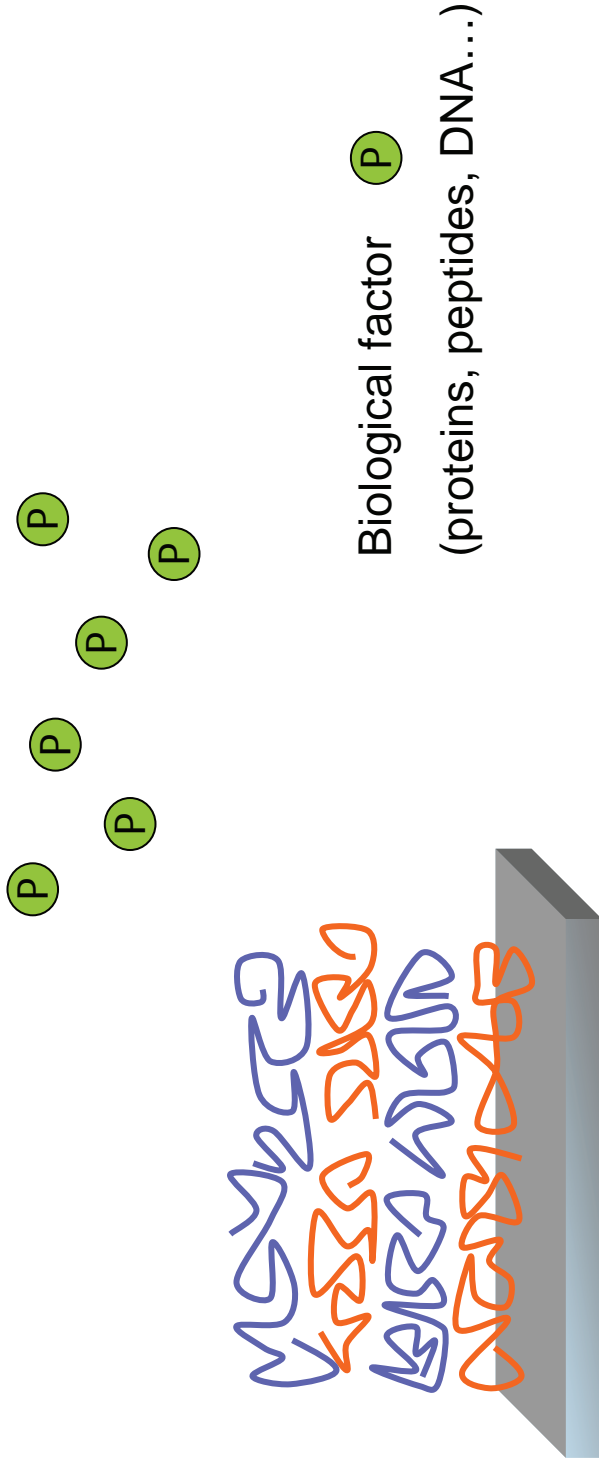
Polyelectrolyte multilayer films as nanoassemblies



1. Polycation solution
2. Buffer
3. Polyanion solution
4. Buffer

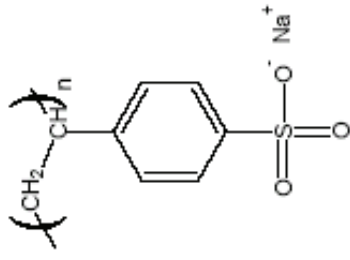


- Films made of biopolymers : polysaccharides, polypeptides
- Confer a biological activity by embedding biological factors

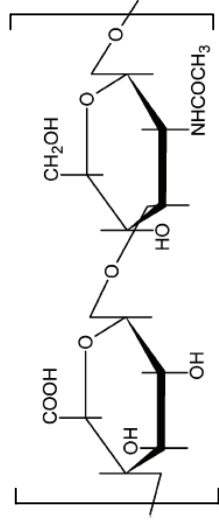


Build up of the Polyelectrolyte multilayers

Polyanions

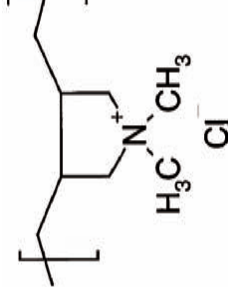


Poly(styrene sulfonate)
PSS

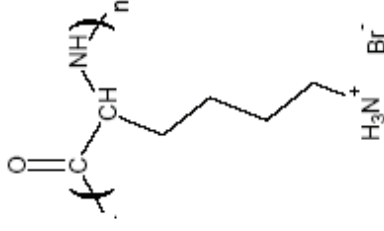


Hyaluronic acid
HA

Polyocations

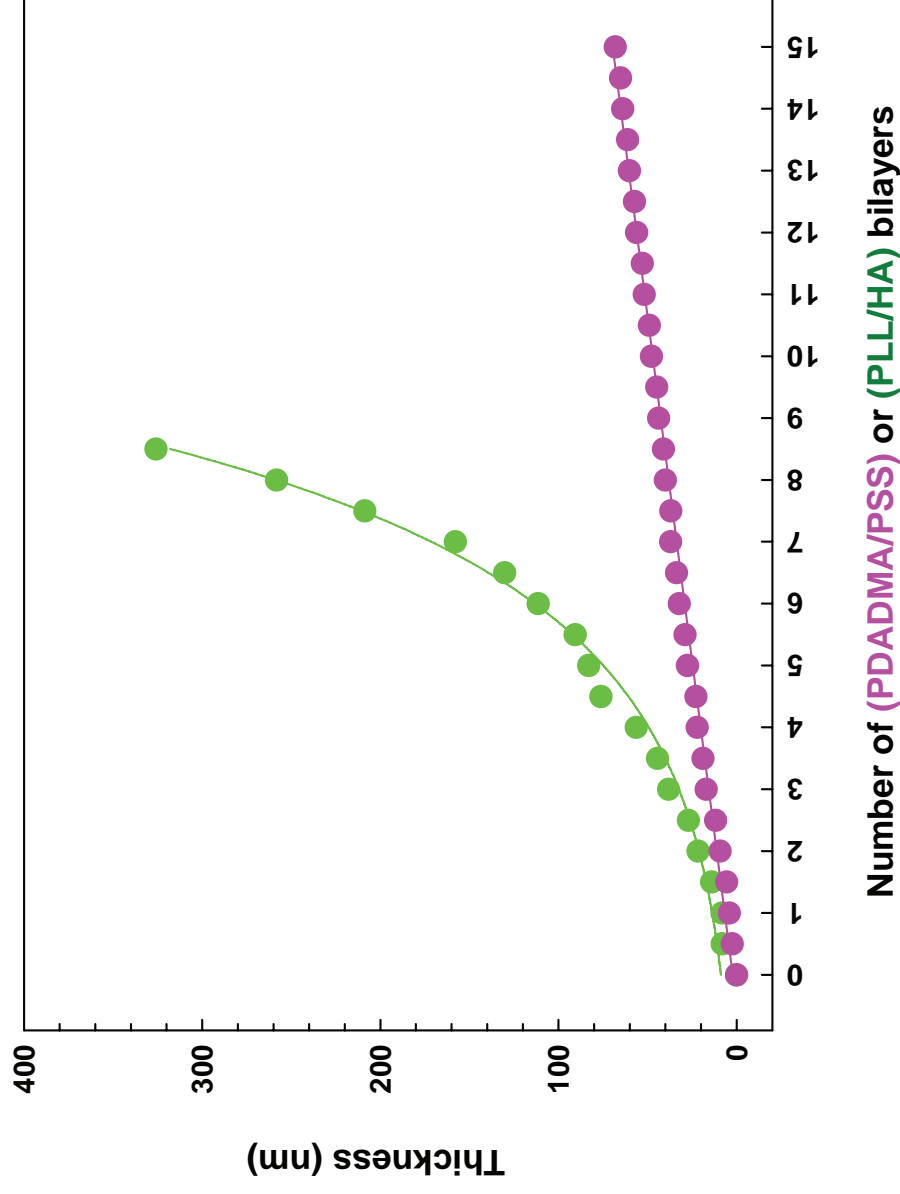


Poly(diallyl-dimethyl-
ammonium chloride)
PDADMA



Poly-L-Lysine
PLL

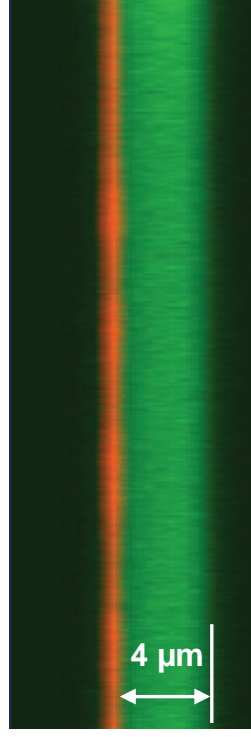
Build up of the Polyelectrolyte multilayers : QCM (Quartz Crystal Microbalance)



Linear growth regime vs Exponential growth regime

 *Macromolecules* 2002; 35: 4458

 *Macromolecules* 2004; 37: 1159



Film section

(PLL / HA)₁₉ / PLL₂₀
with PLL₁₉ FITC and HA₁₉^{TR}

Diffusion of PLL chains through the film section

 *Proc. Natl. Acad. Sci. USA* 2002; 99:12531

 *Langmuir* 2001; 17:7414

PDADMA / PSS vs PLL / HA films

(PDADMA/PSS)

☞ *Linear growth*

☞ *Thin*

☞ *No diffusion through film section*

☞ *Dense*

(PLL/HA)

☞ *Exponential growth*

☞ *Thick*

☞ *Diffusion of PLL through film section*

☞ *Hydrated*

Multilayer Films with compartments / barriers

BARRIERS

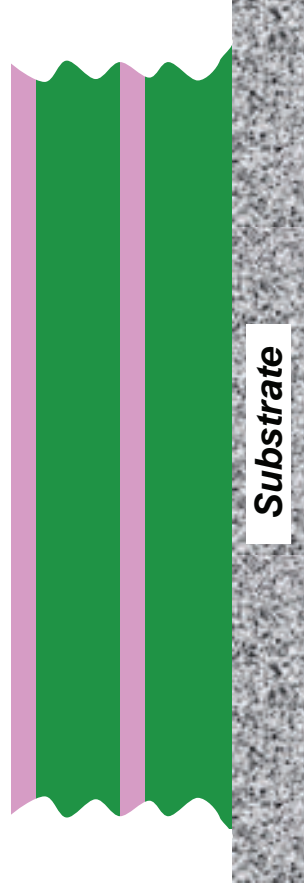
- films with **no diffusion** of polymers, peptides, proteins... through the film section
= *films with linear growth regime like (PAH/PSS) or (PDADMA/PSS) films*

Thin & dense

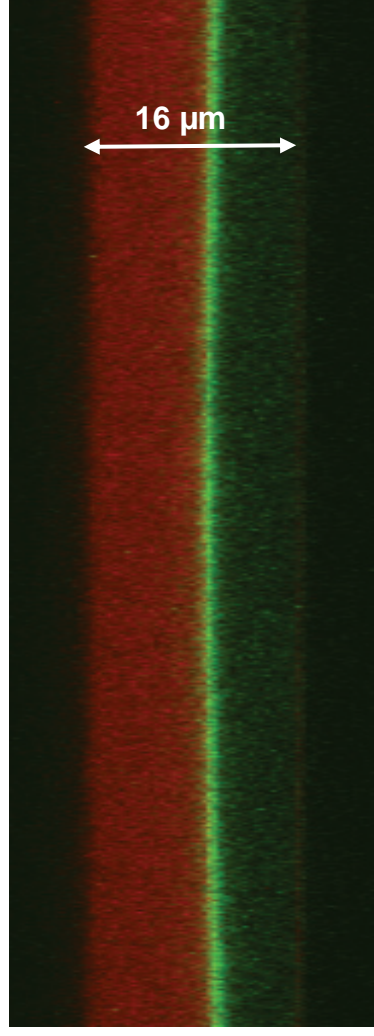
COMPARTMENTS

- films with **diffusion** of polymers, peptides, proteins
= *films with exponential growth regime like (PLL/HA) films*

Thick & highly hydrated



PSS / PDADMA as barrier layers



$(\text{PLL}/\text{HA})_{30}$ / PLL^{FITC} / $(\text{PSS}/\text{PDADMA})$ / HA / $(\text{PLL}/\text{HA})_{30}$ / PLL^{Rho}

📄 *Langmuir* 2004; 20: 7298

📄 *Langmuir* 2005; 21: 12372

📄 *Macromolecules* 2007, 40, 316

Adaptative surfaces made of polyelectrolyte multilayers

 ***Polyelectrolyte multilayer films responding to external stimuli***

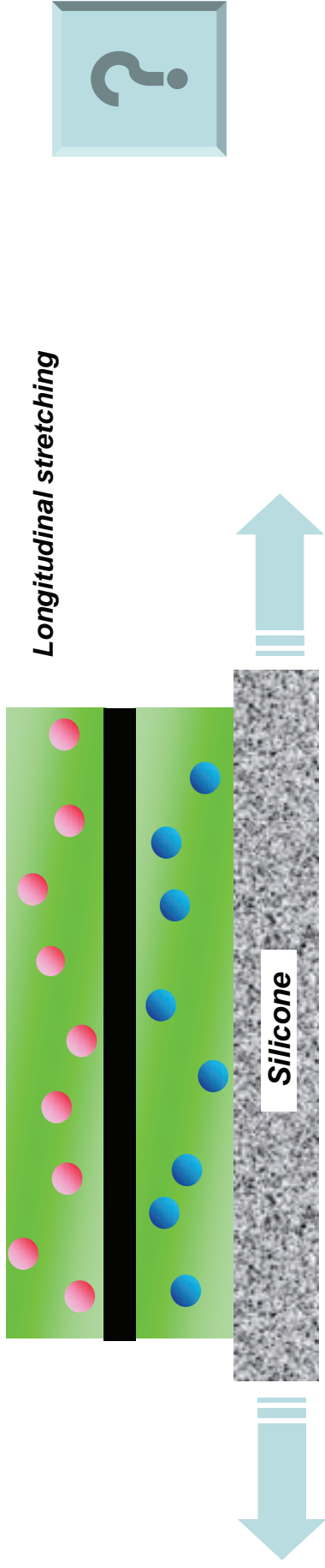
pH
Ionic strength
Temperature

 ***Our objectives :***

To develop coatings responding to *mechanical stimuli*

Applications: Bioactive Patches / Biosensors

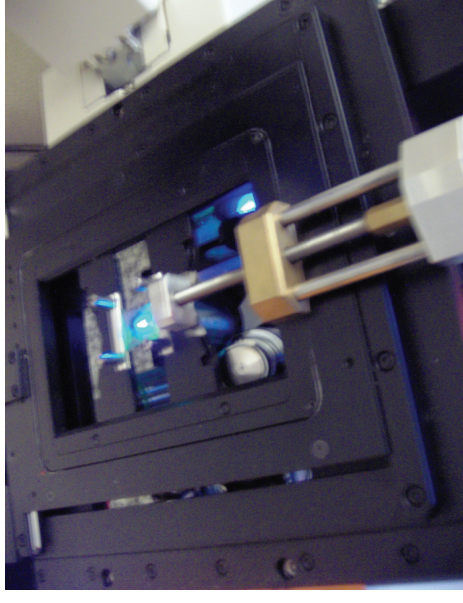
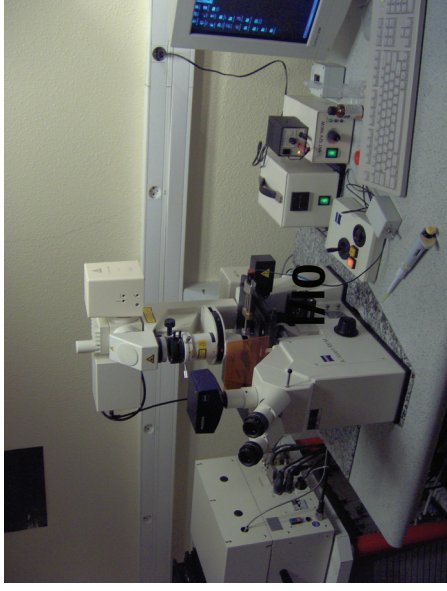
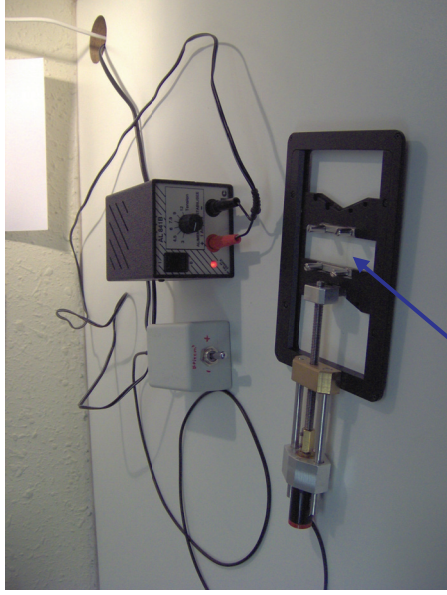
Schedule of conditions



- **No breakage of compartments**
but opening of the barriers under stretching ?
- **Mixing 2 components to trigger enzymatic or chemical reactions,**
Release of active compounds under stretching

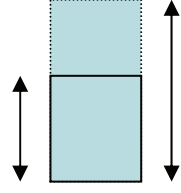
Methods

Homemade stretching devices adapted to AFM, CLSM... techniques



Stretching degree is defined by the elongation parameter $\alpha = l/l_0$

l_0 initial length



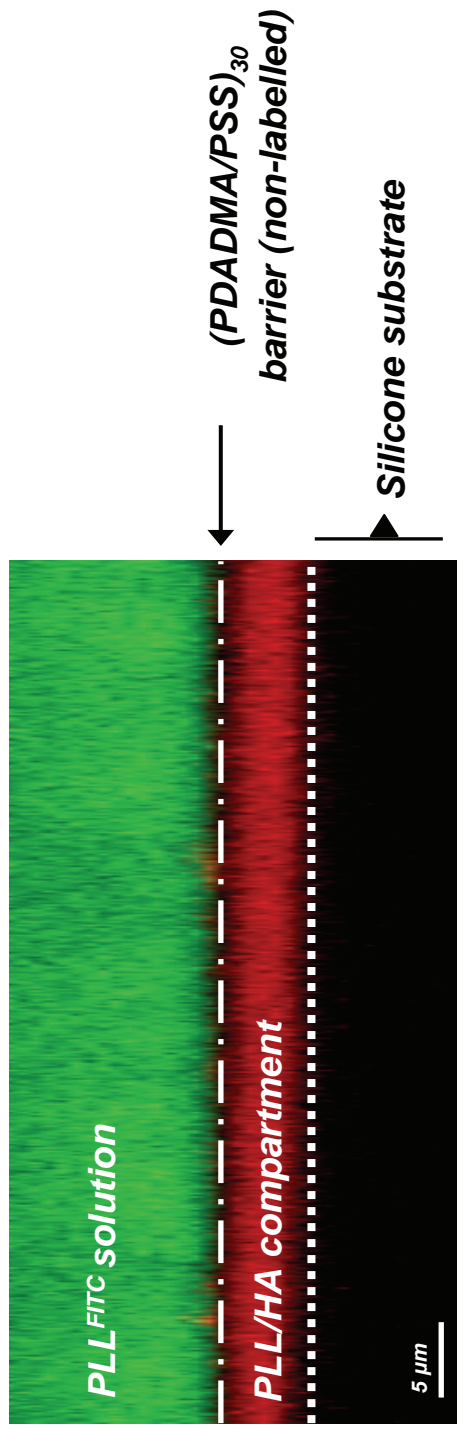
l stretched length

- $\alpha = 1$ No stretching
- $\alpha = 2$ Elongation of the silicone sheet X2

Compartment / Barrier systems

(PLL/HA)₃₀ labelled in red - (PDADMA/PSS)_n capping
+ **PLL^{FITC}** green solution at the non-stretched state

Confocal (x,z) section in liquid condition

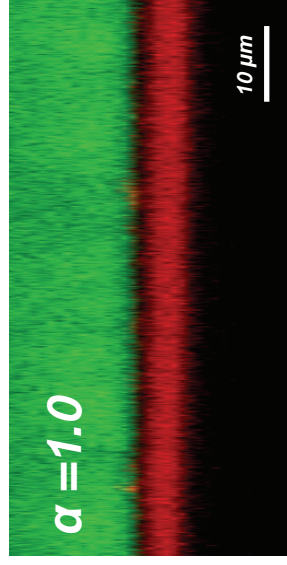


No green color detected in the compartment over more than several hours

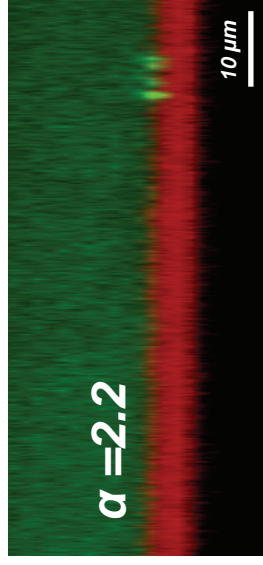
Compartment / Barrier systems

$(PLL/HA)_{30}$ labelled in red - $(PDADMA/PSS)_n$ capping + PLL^{FITC} solution under stretching

$n=30$

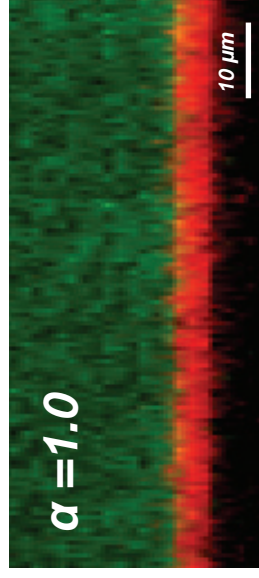


$\alpha = 2.2$

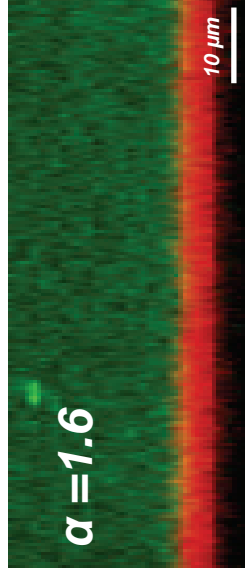


Confocal x,z sections
in liquid conditions

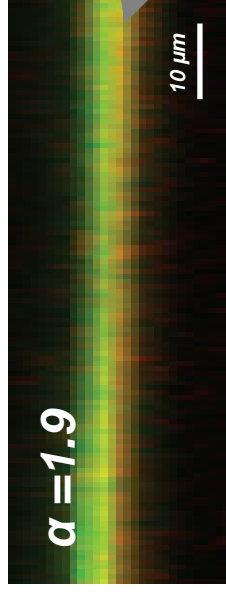
$n=15$



$\alpha = 1.6$

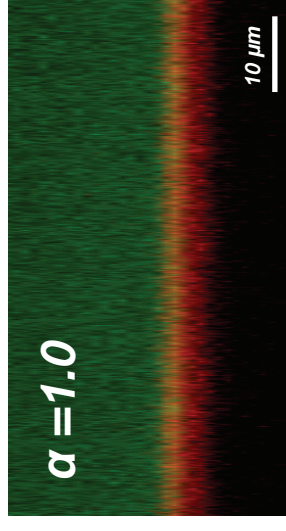


$\alpha = 1.9$

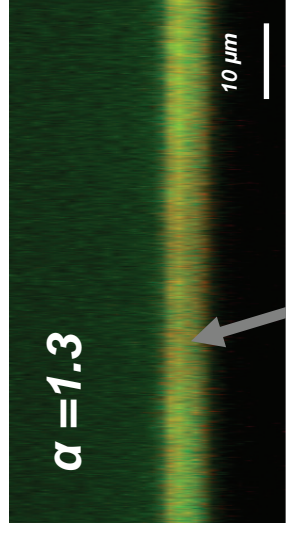


$n=5$

$\alpha = 1.0$

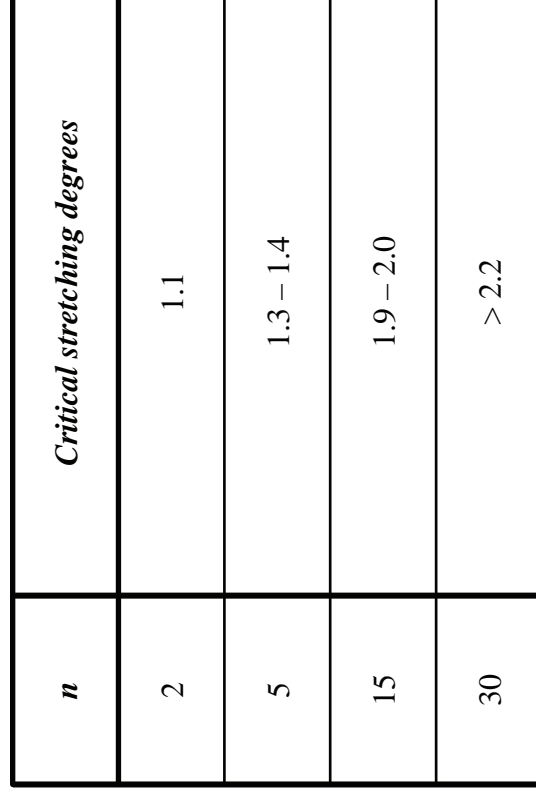


$\alpha = 1.3$



PLL^{FITC} diffusion from the
solution to the compartment

Influence of n upon the critical stretching degree



n	Critical stretching degrees
2	1.1
5	1.3 – 1.4
15	1.9 – 2.0
30	> 2.2

increasing with n

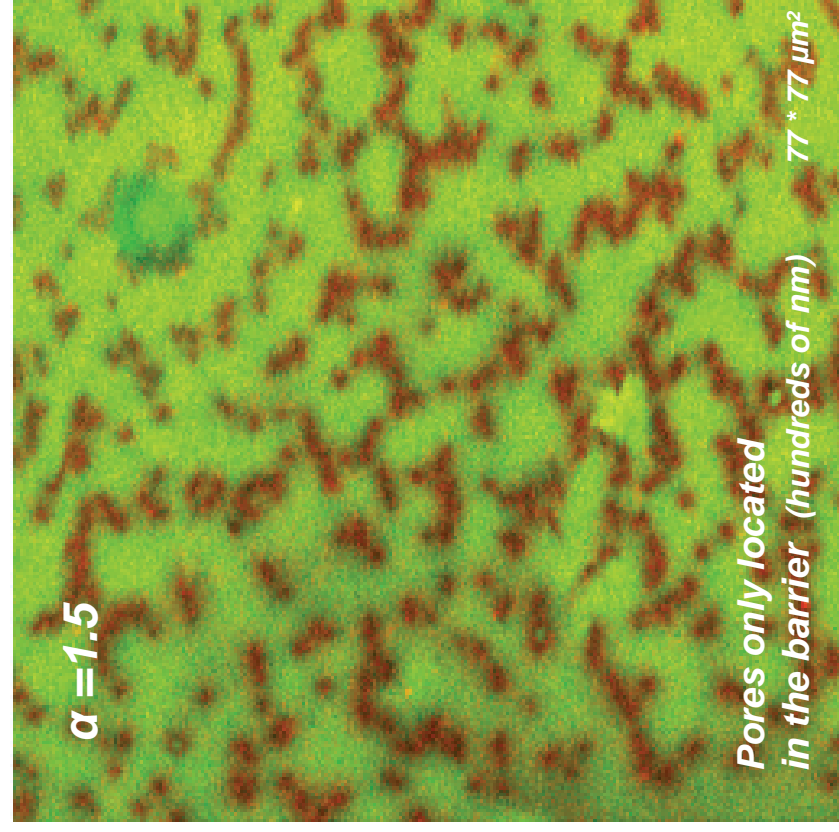


Tune the response by adjusting n

Formation of nanopores in the barrier

CLSM (x,y) images at the film/solution interface

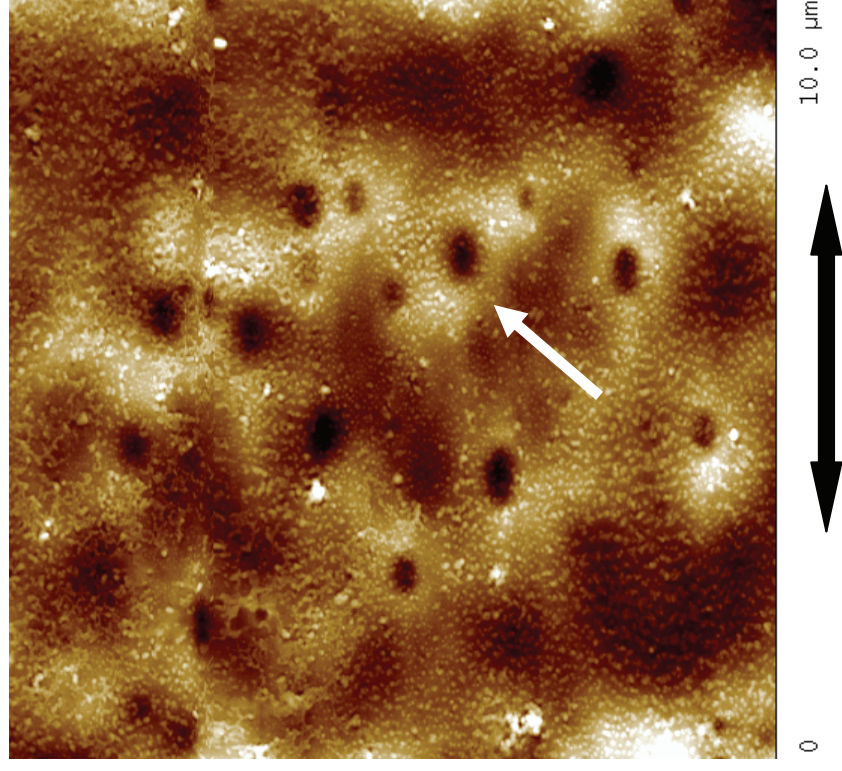
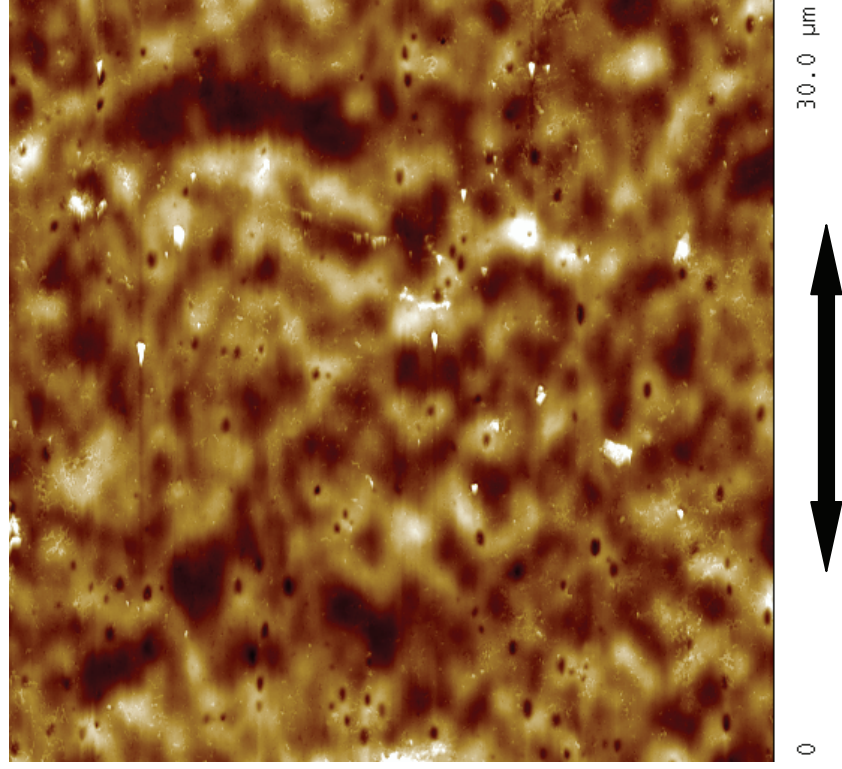
(PLL/HA)₃₀ labelled in red - (PDADMA/PSS)₅ barrier + **PLL^{FITC}** solution



Formation of nanopores in the barrier

AFM images under mechanical stretching at 2.0

$(PLL/HA)_{30}$ compartment - $(PDADMA/PSS)_5$ barrier



Pore sizes in accordance with CLSM measurements

Two-compartment systems

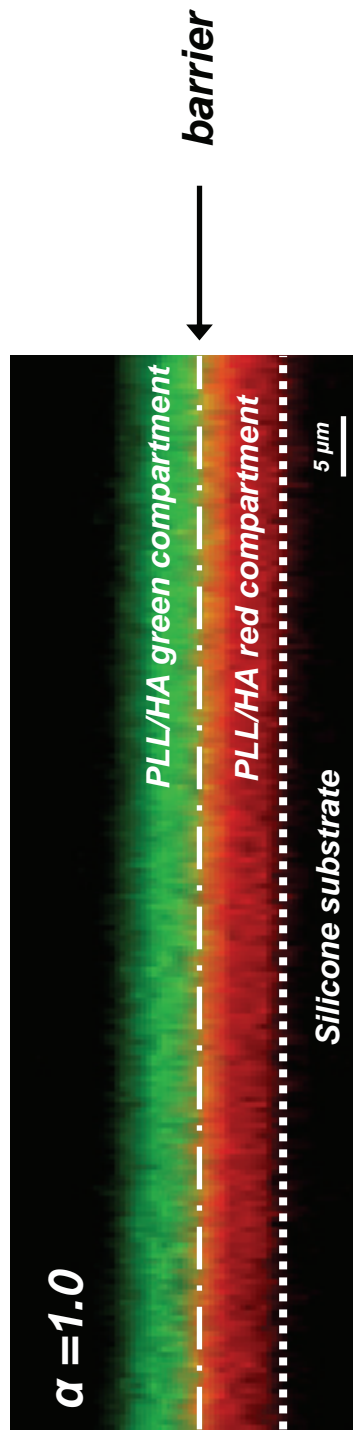
Compartment/Barrier/Compartment systems

$n=5$

(PLL/HA)₃₀ compartment labelled in red

(PDADMA/PSS)₅ barrier

(PLL/HA)₃₀ compartment labelled in green



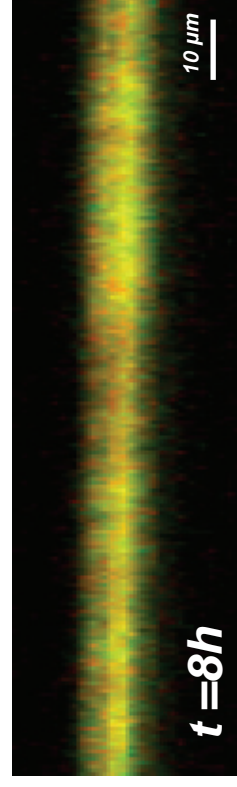
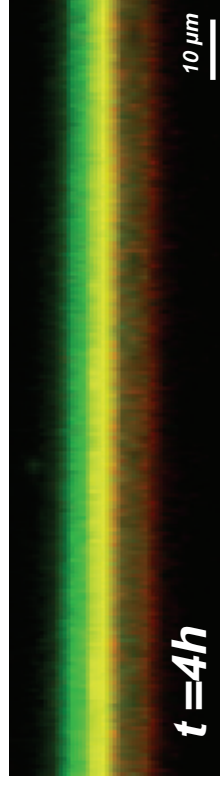
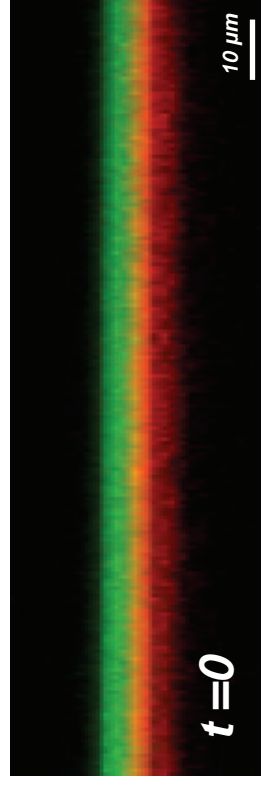
➤ **No diffusion of PLL chains through the barrier**

Two-compartment systems

Control of PLL mixing by mechanical stretching, $\alpha = 1.9$

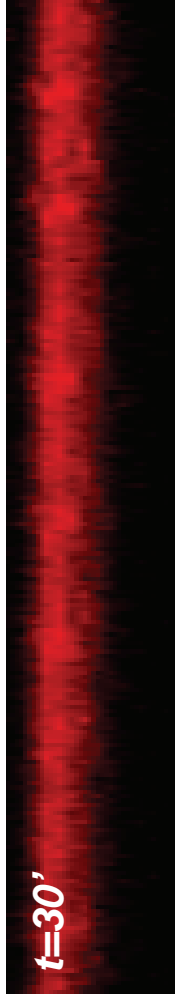
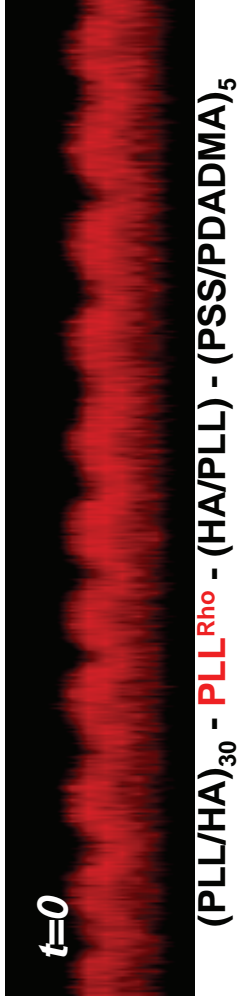
Time evolution under stretching for :

$(\text{PLL/HA})_{30}$ labelled in red - $(\text{PSS/PDADMA})_5$ - $(\text{PLL/HA})_{30}$ labelled in green

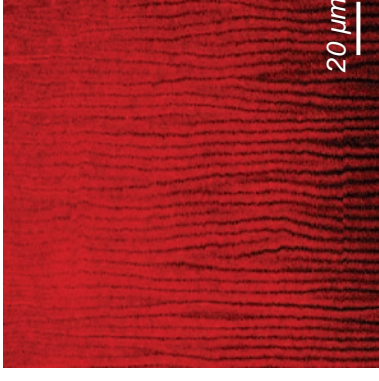
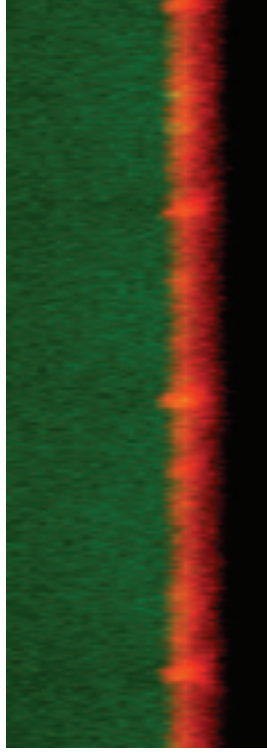


Reversibility of the mechanism : PSS/PDADMA wound healing

Back to $\alpha = 1.0$

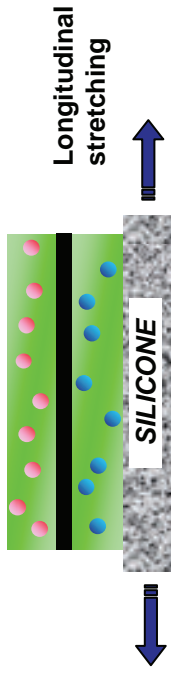


+ PLL^{FITC}



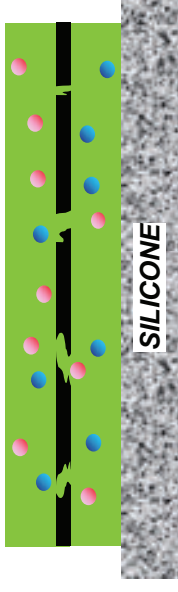
Barrier becomes again efficient
and prevents PLL diffusion,
pores are closed

Results

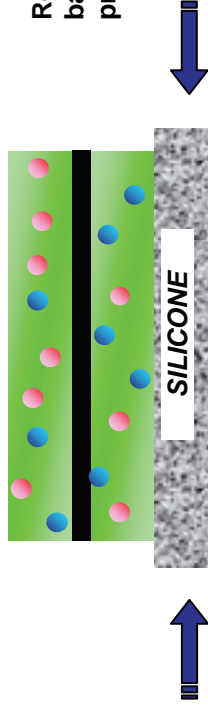


Longitudinal stretching

Multicompartments films under mechanical stretching



Nanopores in the barrier allow a diffusion process



Return to the unstretched state: barrier closing by « healing » process

Pore formation is a reversible process : nanovalves

Potential Applications :

biocatalytic surfaces with enzymatic activity controlled by mechanical stretching

 *Nano Letters*, 7, 657, 2007.

 *Soft Matter*, 3, 1413, 2007.

Acknowledgements

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