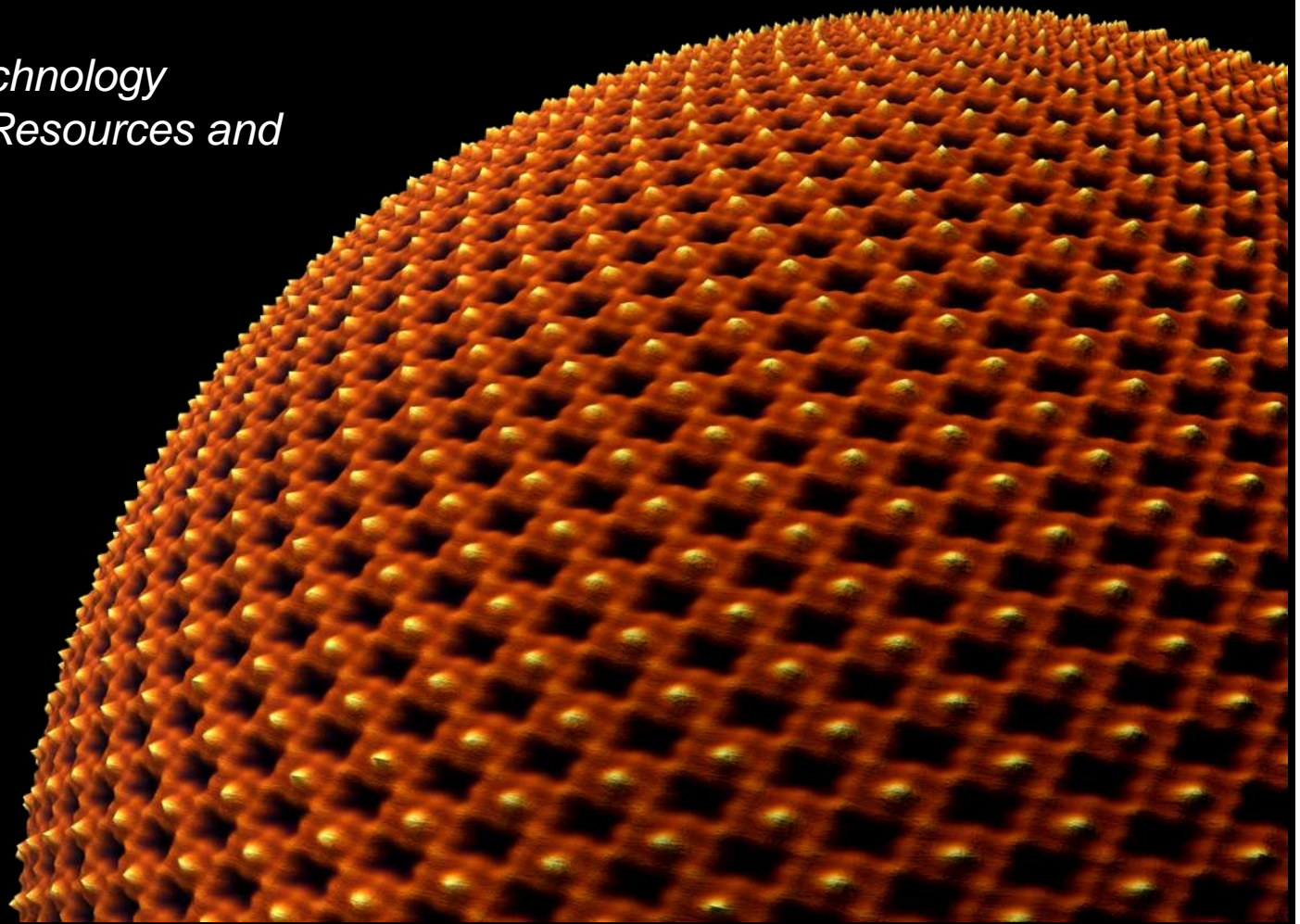


# Functionalization of surfaces with S-layer protein lattices

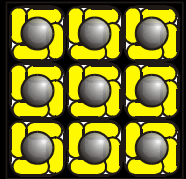
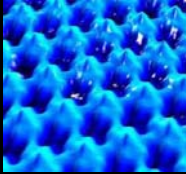
*Dietmar Pum, Bernhard Schuster, Nicola Ilk, and Uwe B. Sleytr*

*Center for NanoBiotechnology  
University of Natural Resources and  
Applied Life Sciences  
Vienna, Austria*

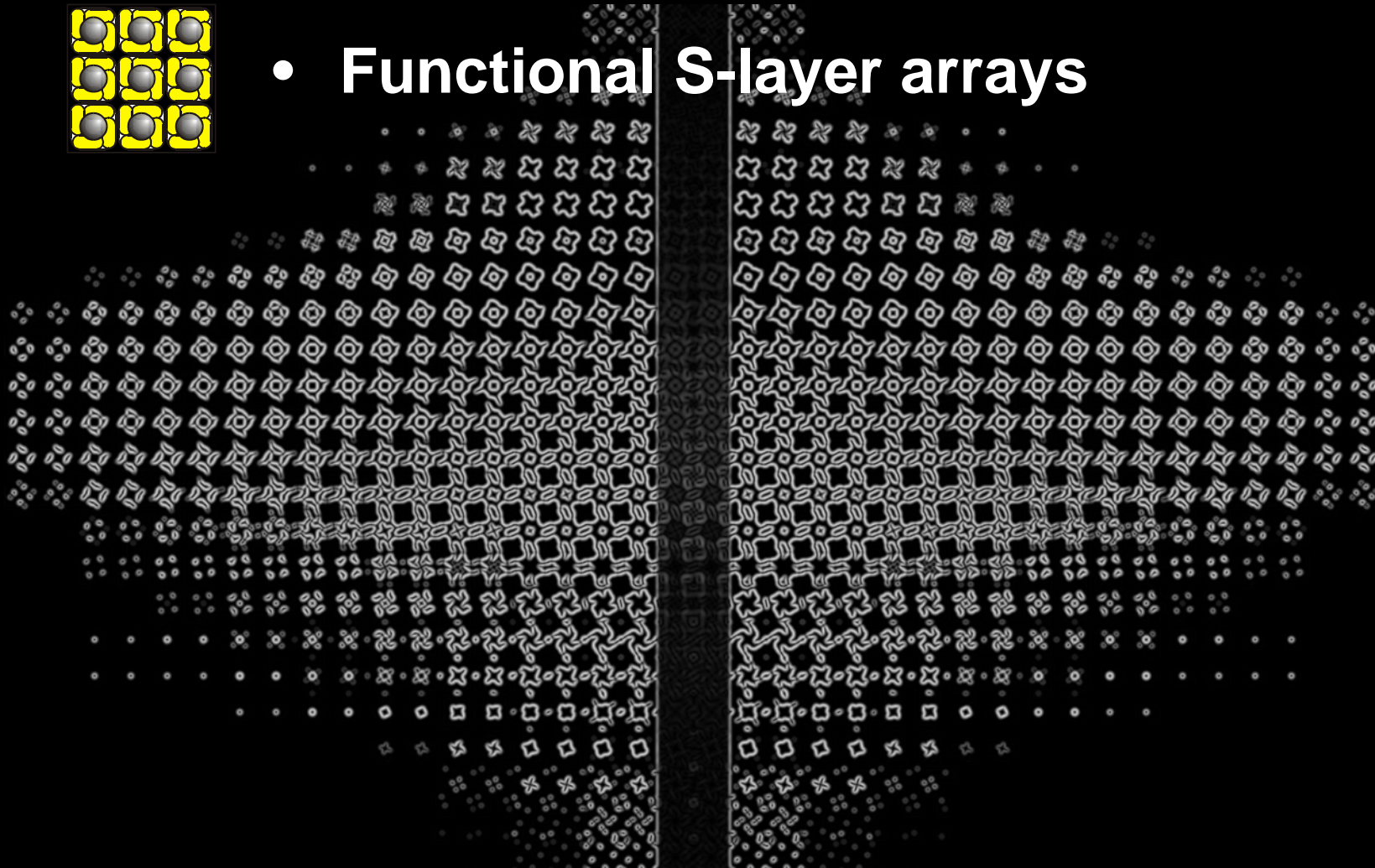


NanoBioEuro 2008  
Barcelona

## Outline:



- Description of S-layer proteins
- Functional S-layer arrays





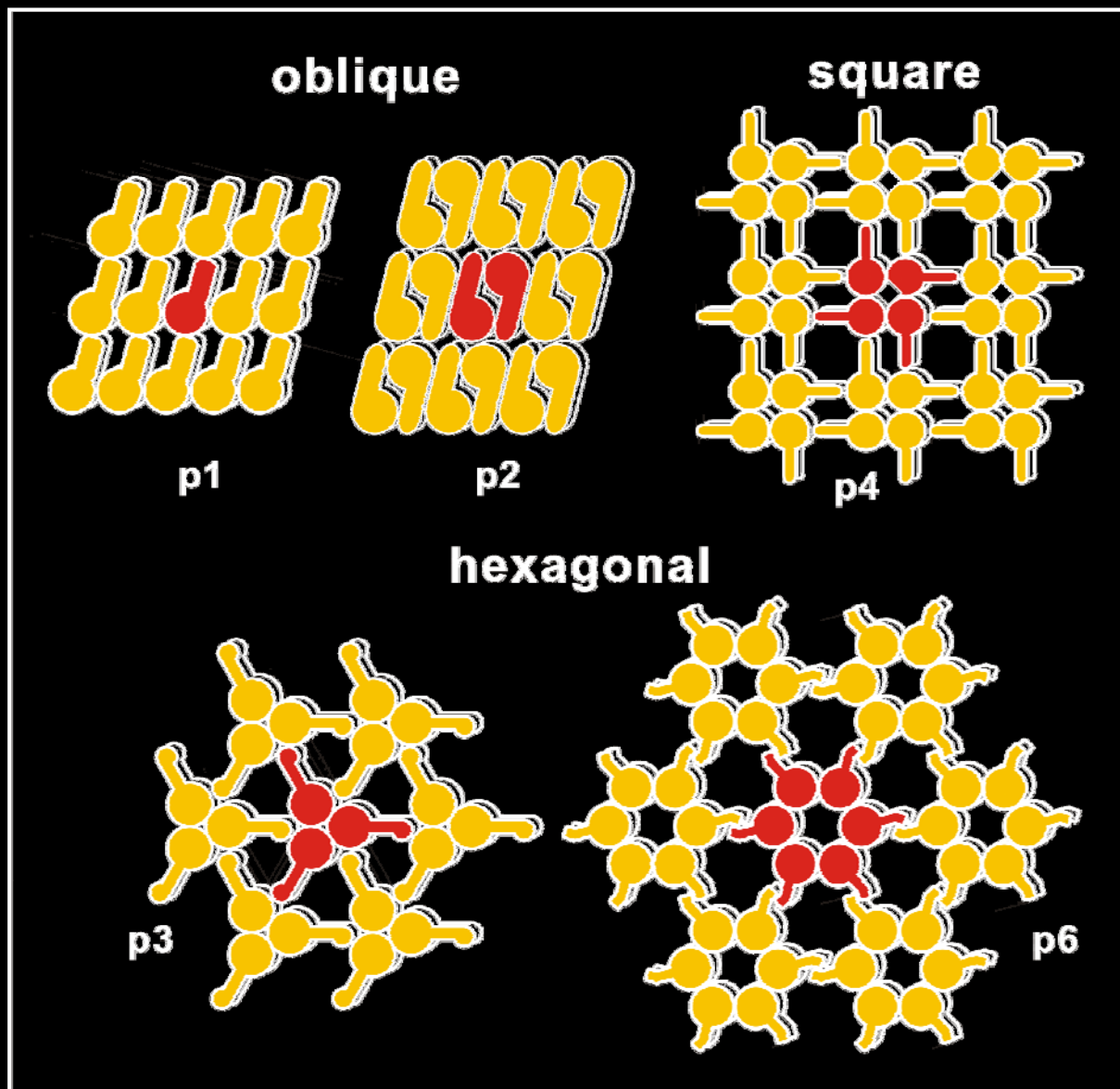
# Description of S-layers



Crystalline bacterial cell surface layer proteins (**S-layer proteins**) represent the outermost cell envelope component in a broad range of eubacteria and archaea.

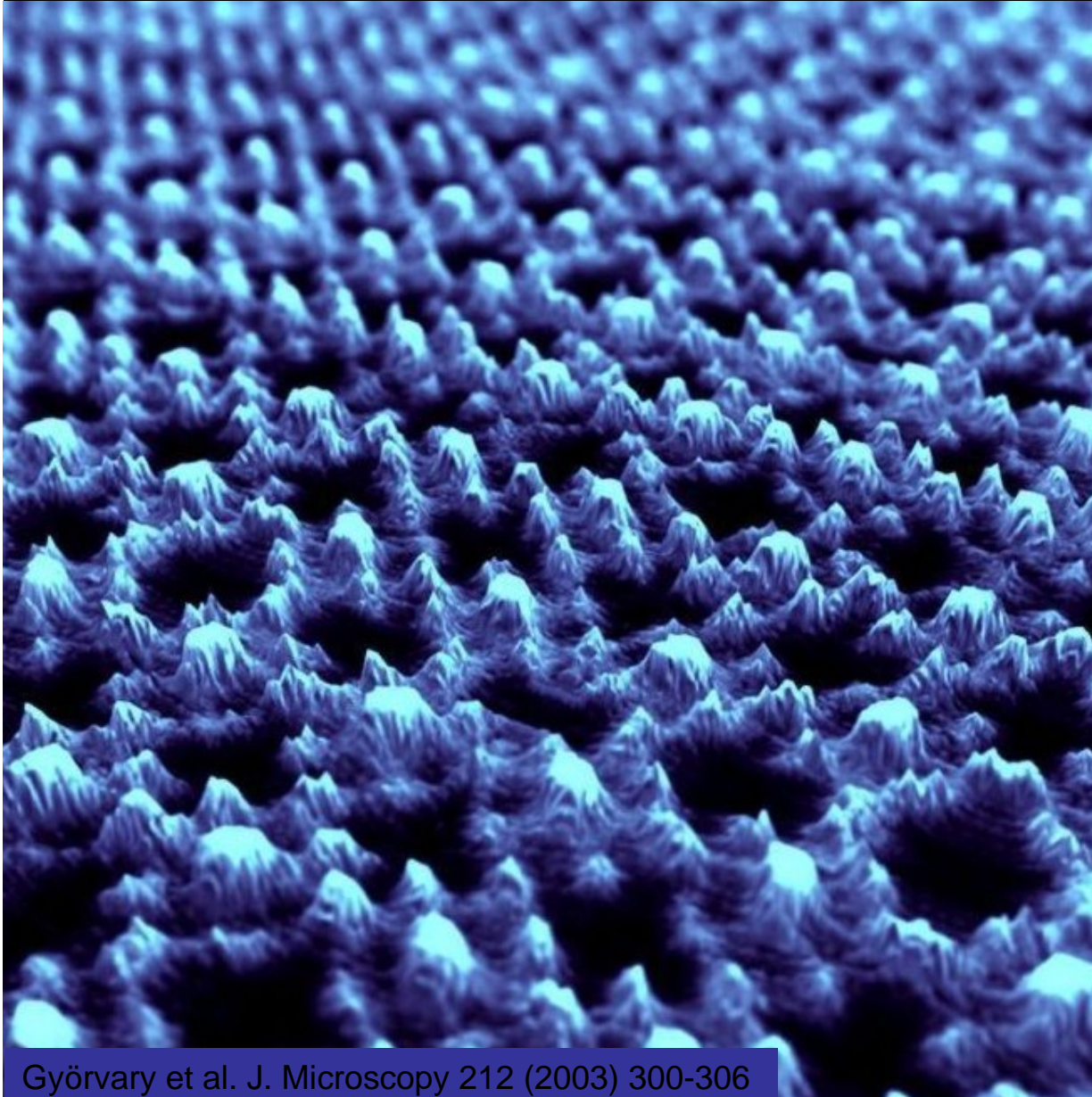
TEM of bacterial cells with an S-layer showing square lattice symmetry.

# S-layer lattice types





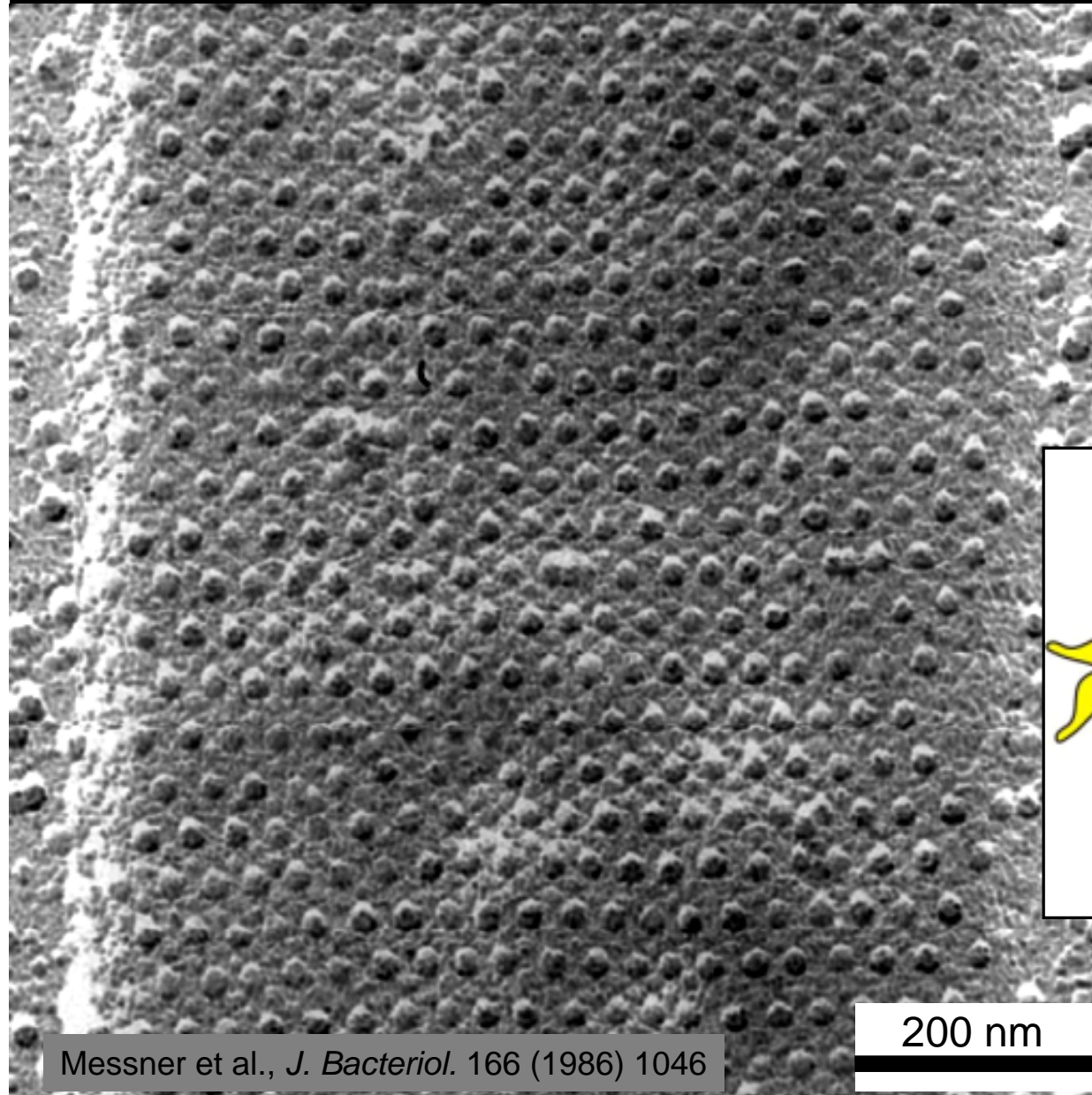
# Ultrastructure of S-layers



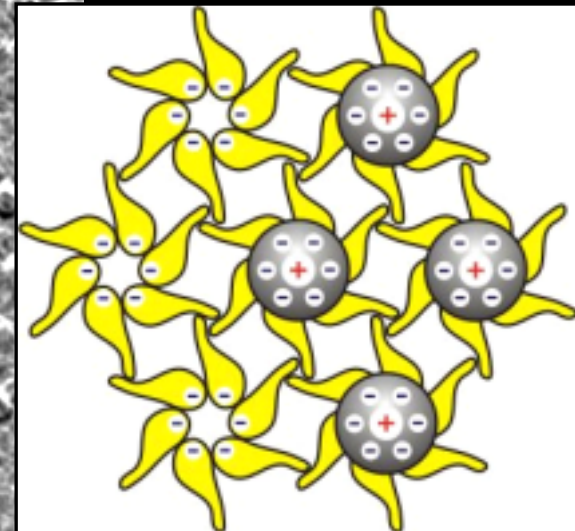
- $M_r$ : 40-200 kDa
- Unit cell sizes:  
3 – 30 nm
- Thickness:  
5 – 10 nm
- Pores are of  
identical size and  
morphology

AFM image of an S-layer showing  
square lattice symmetry ( $d=13.1\text{nm}$ ).

# S-layers as patterning elements



Functional groups and domains are repeated with the periodicity of the lattice

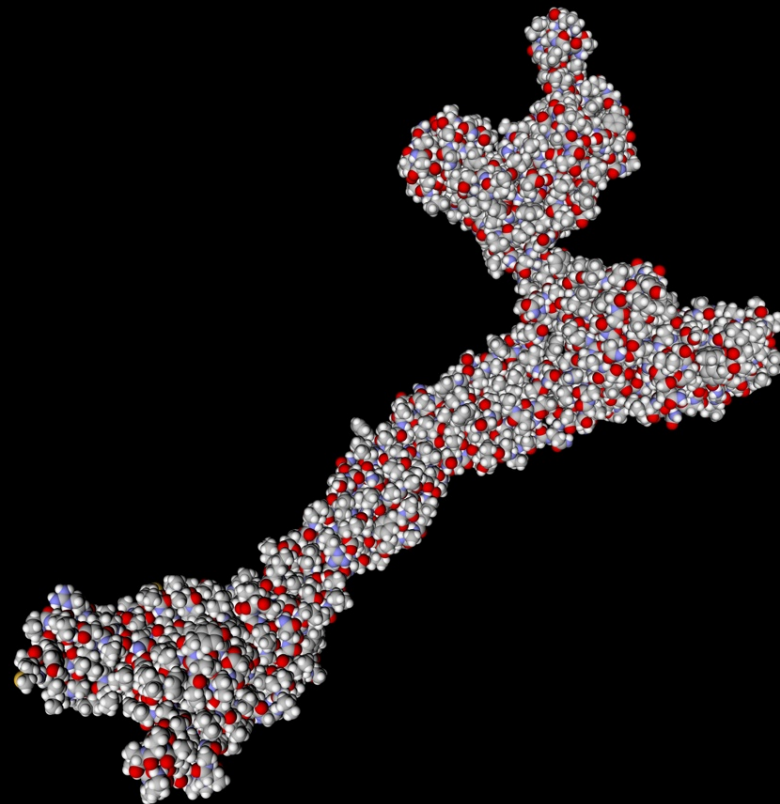
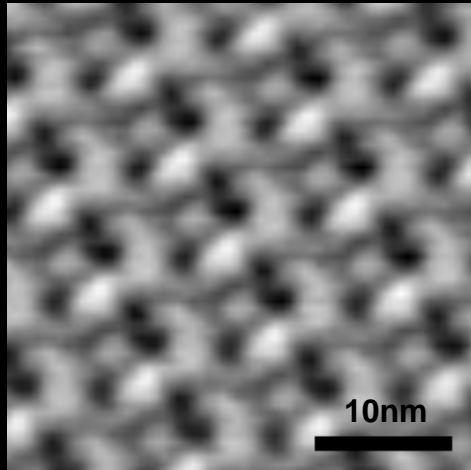


TEM image of PCF molecules bound to an S-layer showing hexagonal lattice symmetry.



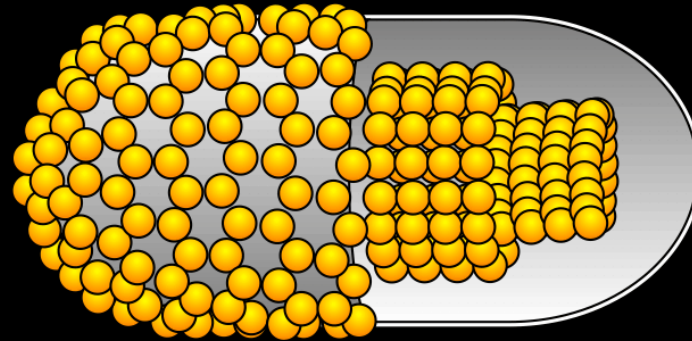
# 3D-structure prediction by molecular modeling

(S-layer protein SbsB from *Geobacillus stearothermophilus* pv72/p2)



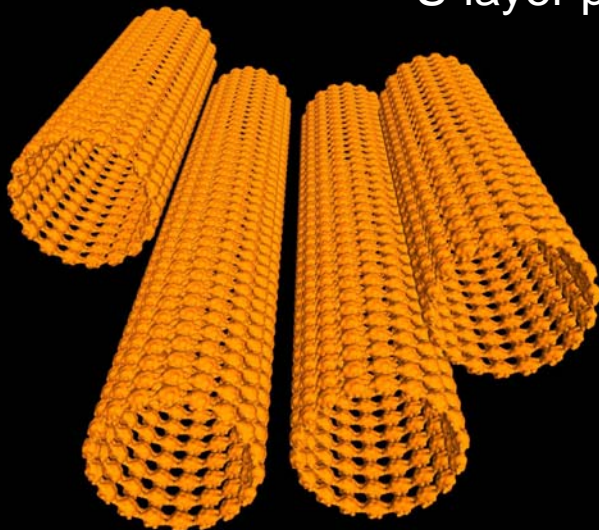
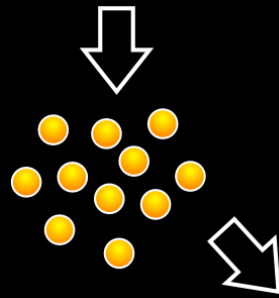
# Reassembly of S-layer proteins

bacterial cell

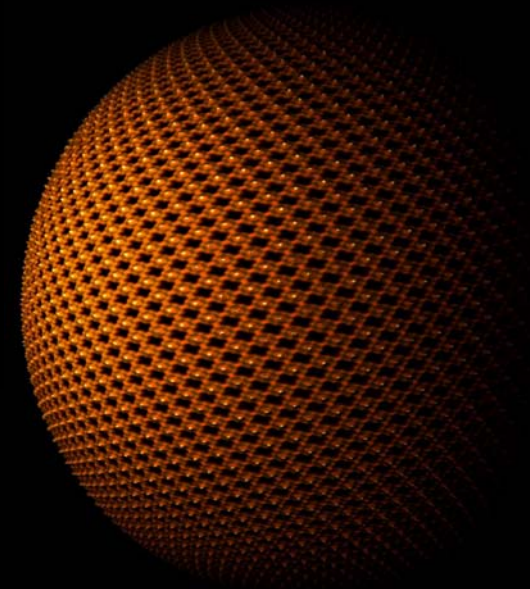
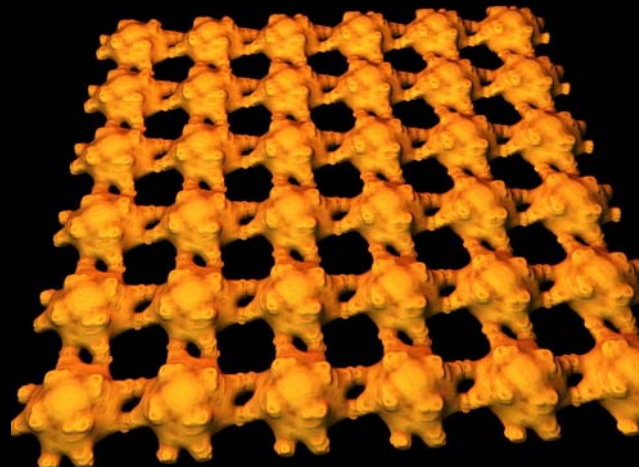


expression host  
(e.g. *E.coli*)

reassembly of native and  
recombinantly produced  
S-layer proteins



cylindrical self-assembly  
products in solution

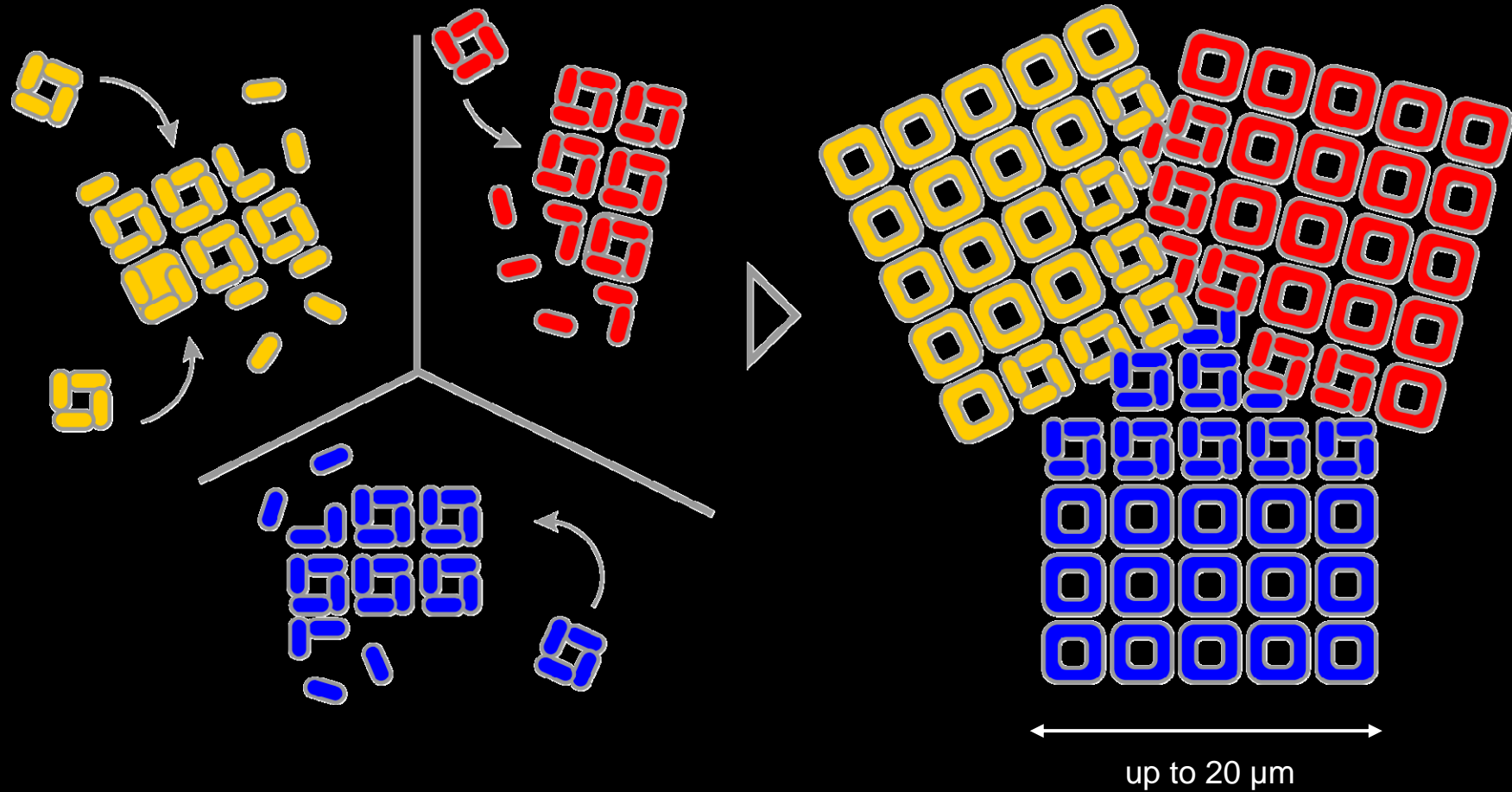


planar lipid membranes,  
liposomes and nanocapsules

monolayers in solution  
and at interfaces



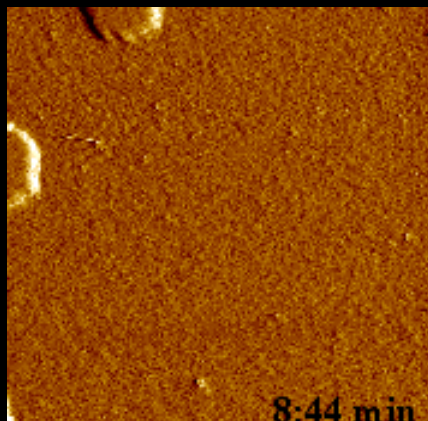
# Crystal growth at interfaces



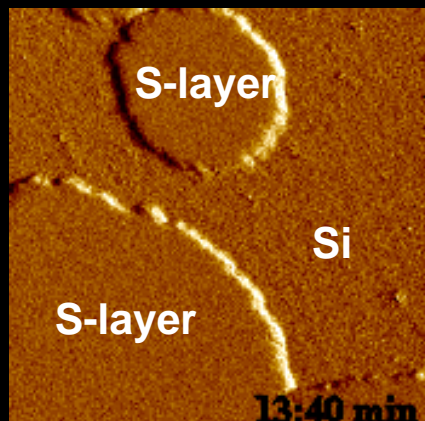
Crystal growth of S-layers on solid supports, at the liquid-air interface or on lipid films.

# Crystal growth on silicon

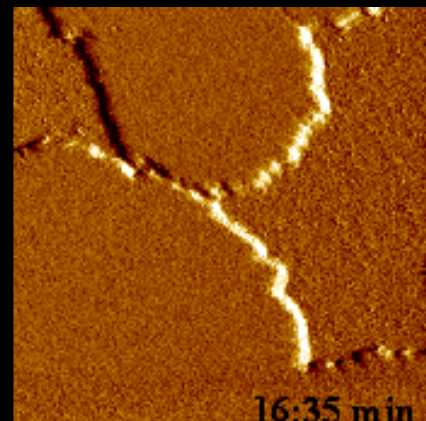
5x5  $\mu\text{m}$



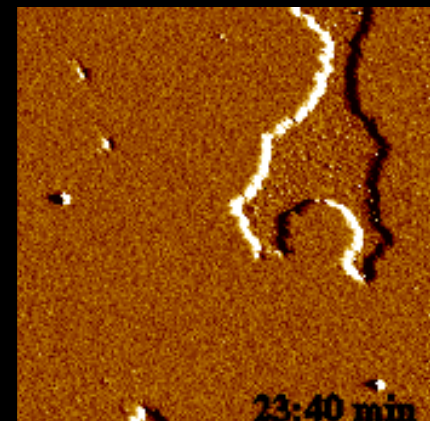
5x5  $\mu\text{m}$



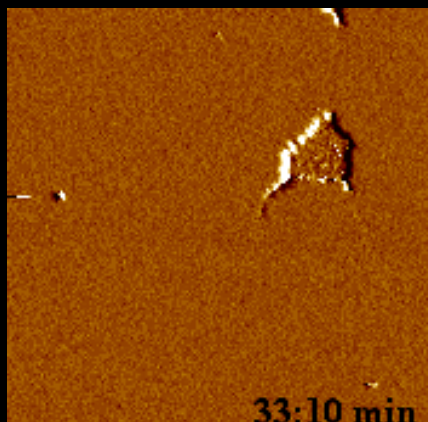
5x5  $\mu\text{m}$



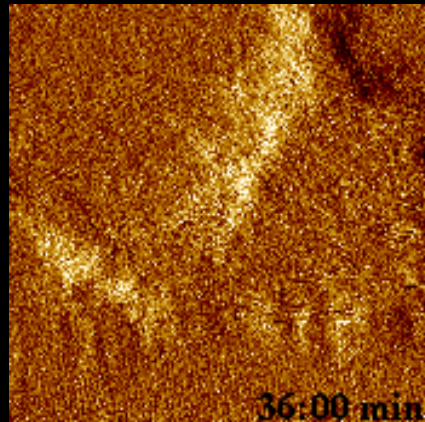
10x10  $\mu\text{m}$



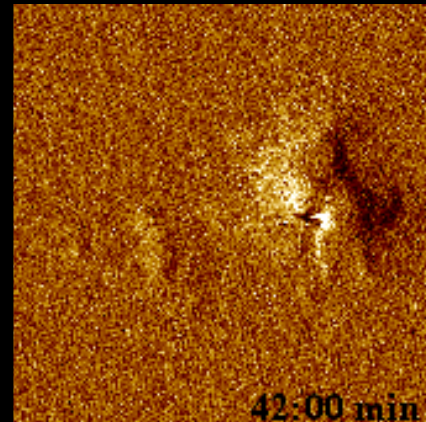
10x10  $\mu\text{m}$



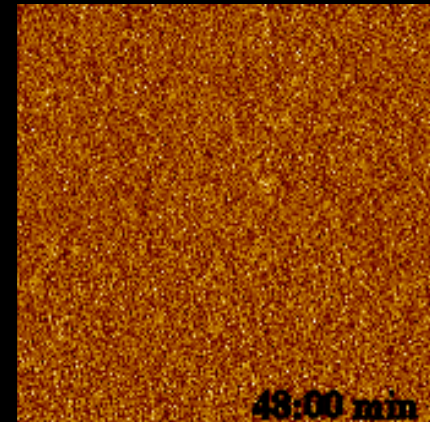
1x1  $\mu\text{m}$



1x1  $\mu\text{m}$

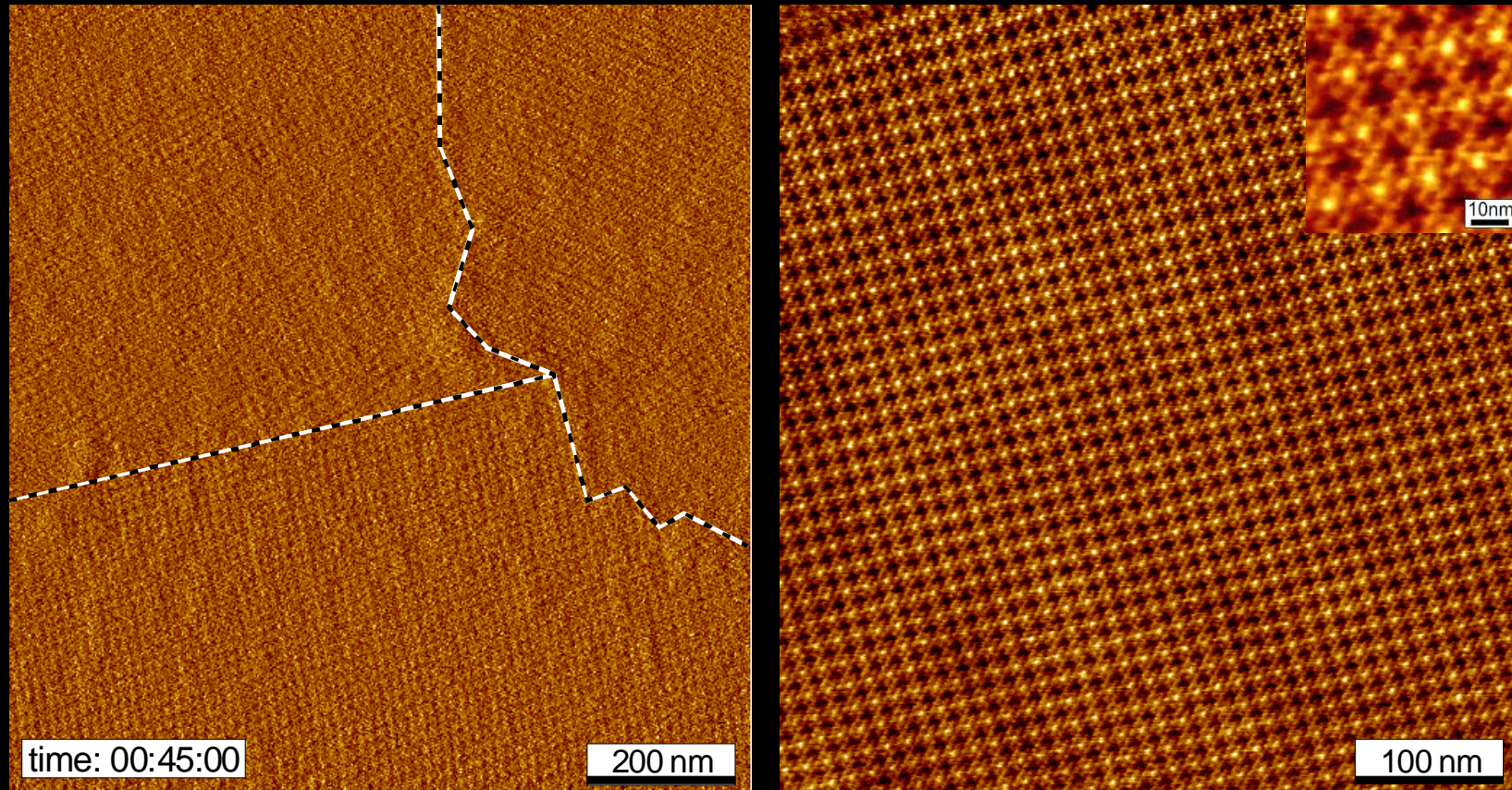


1x1  $\mu\text{m}$



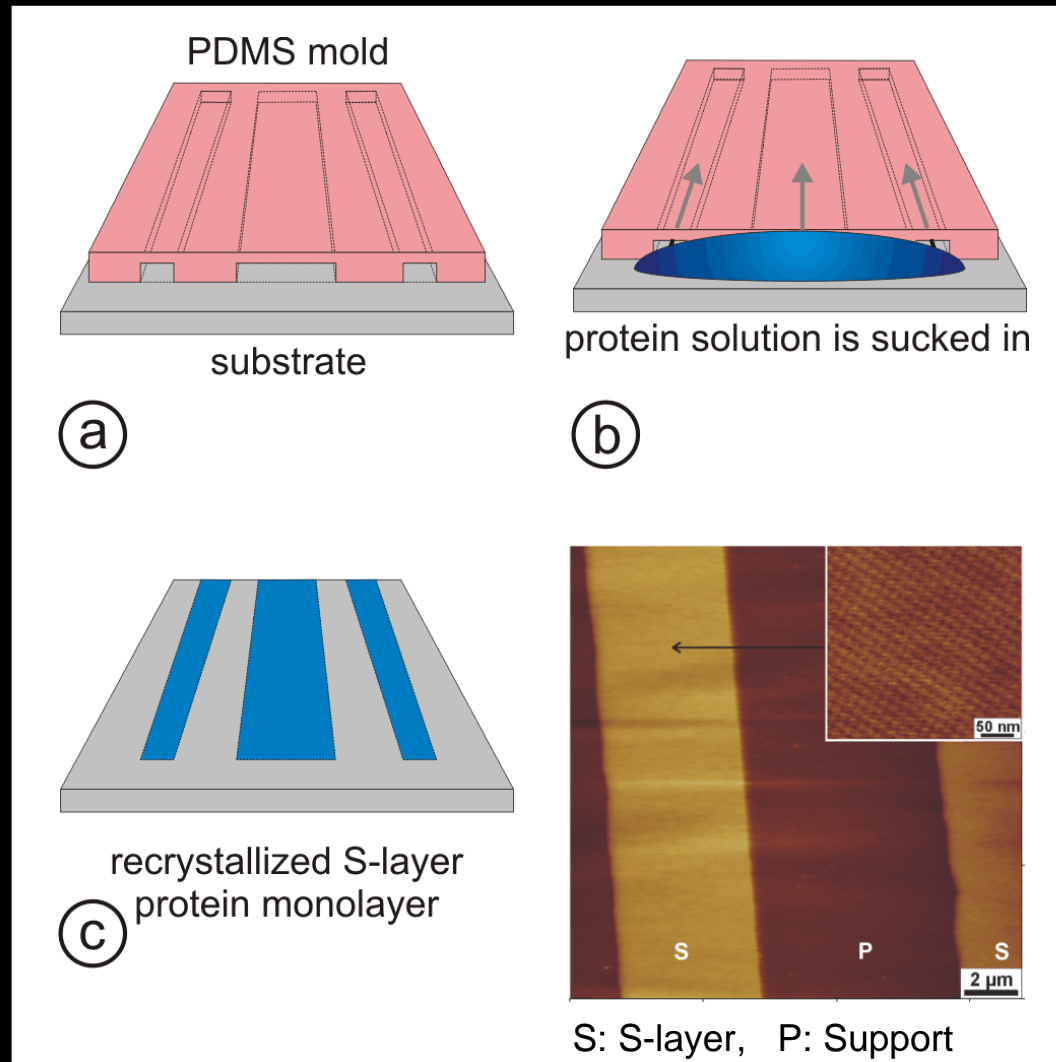


## Formation of an S-layer protein monolayer on silicon



AFM images of the recrystallization process of SbpA S-layer proteins from *B. sphaericus* CCM2177 on silicon

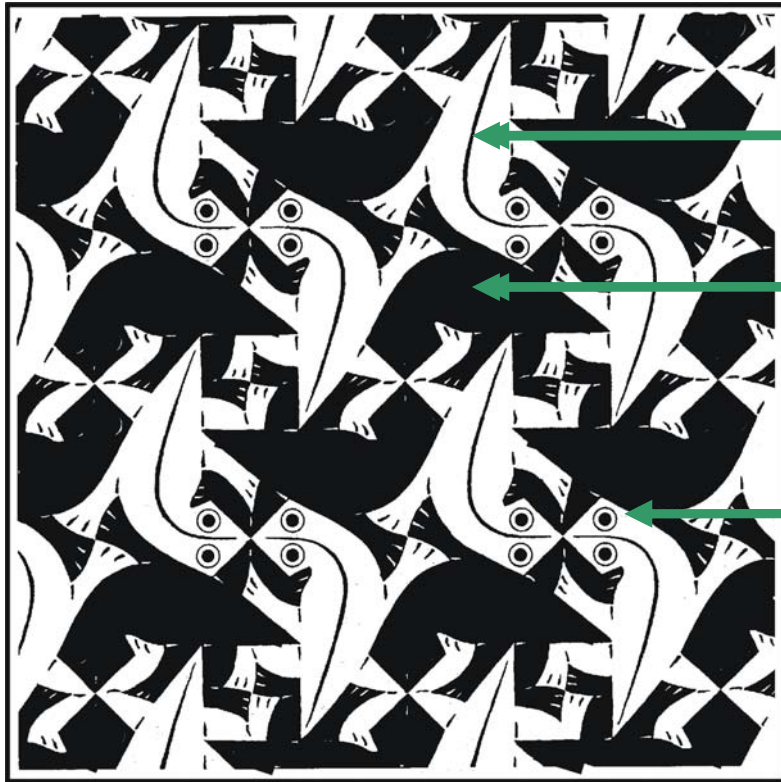
# Patterning of S-layers by Soft Lithography (Micromoulding in Capillaries - MIMIC)





# Functional S-layer arrays

- S-layer functionalized solid supports
- S-layer supported lipid membranes
- S-layer coated liposomes



S-layer lattices are composed of identical species of subunits.

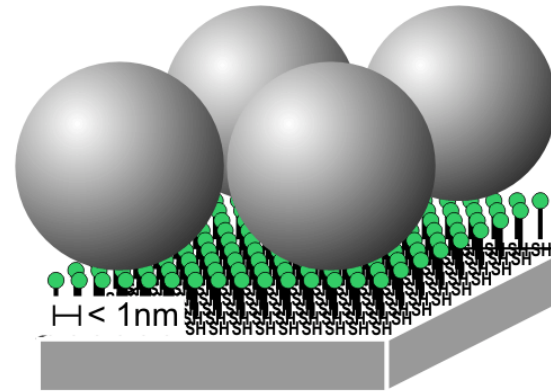
Pores passing through show identical size and morphology.

Functional groups are aligned in well defined positions and show defined orientations.

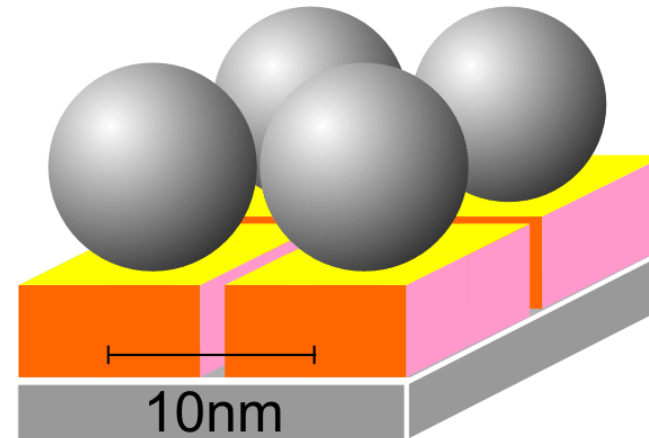
H.C. Escher

# Strategies for modification and functionalization of solid supports

- Self-assembled monolayers (SAMs)

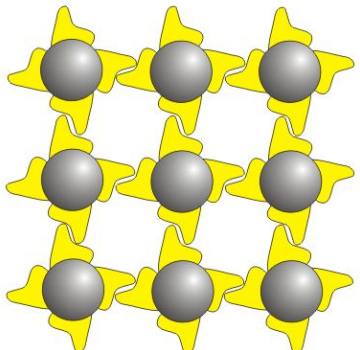
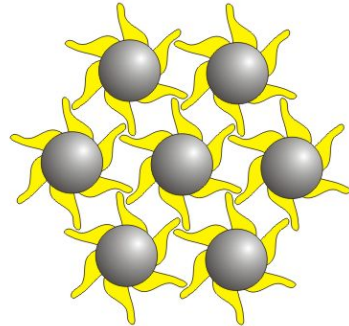
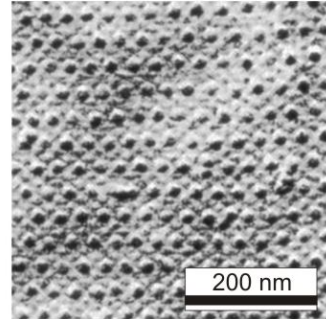
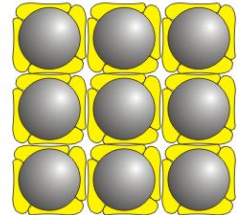
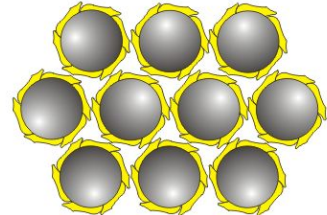
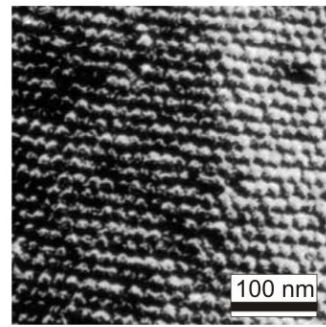


- S-layers lattices





# Binding of macromolecules and nanoparticles into regular arrays on S-layers

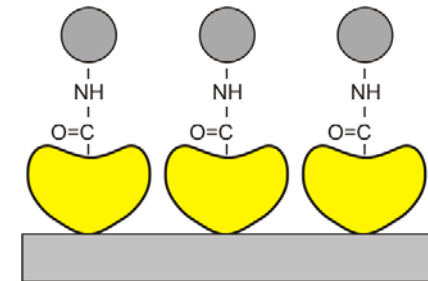
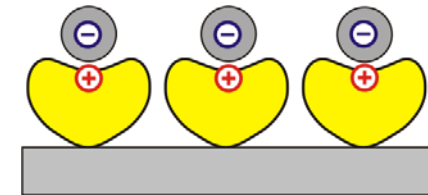
	Lattice symmetry		Examples
Lattice spacing	 square (p4)	 hexagonal (p6)	
	 square (p4)	 hexagonal (p6)	

The superlattice of bound molecules and nanoparticle resembles the lattice parameters of the underlying S-layer

# Binding of molecules and nanoparticles on S-layers

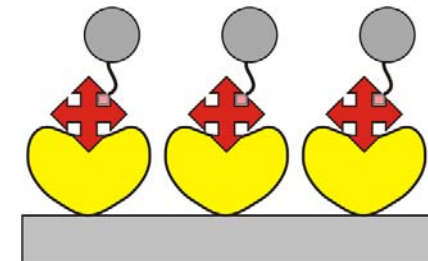
## Native S-layer protein:

- Binding by surface chemical methods using electrostatic bonds (e.g. free carboxyl groups) covalent bonds ( e.g. EDC-activated carboxyl groups )



## Genetically engineered S-layer protein:


- Design and expression of recombinant S-layer fusion proteins.
- Binding by functional domains on S-layer fusion proteins



Functional domains (native or genetically introduced) are repeated with the periodicity of the S-layer lattice leading to regular arrays of bound molecules and particles.

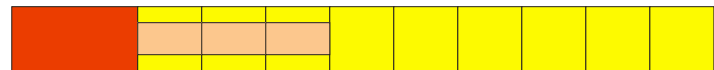
# Binding of biotinylated molecules and nanoparticles on S-layer streptavidin fusion proteins

"C-terminal fusion proteins (BS1)"

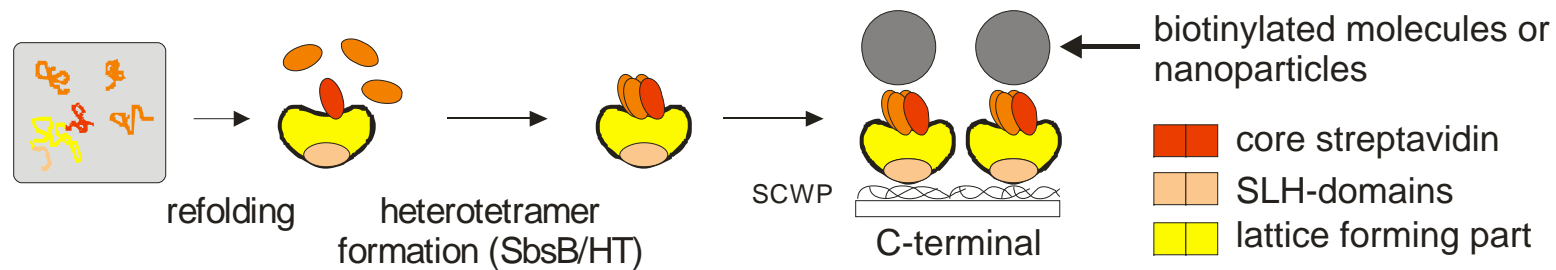


aa sequence

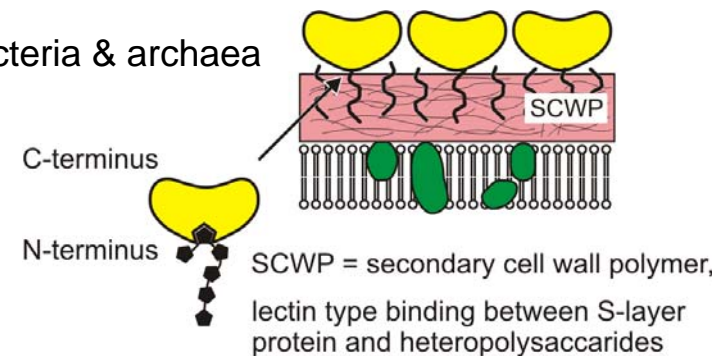
"N-terminal fusion proteins (S1B32)"



aa sequence



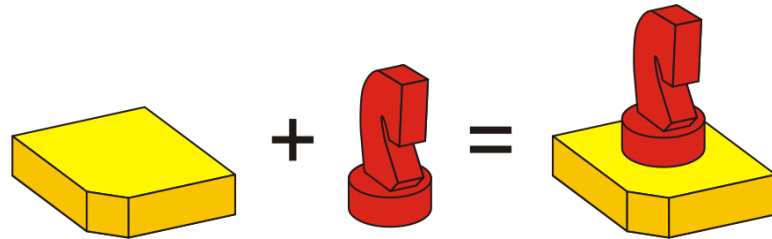
Gram positive bacteria & archaea



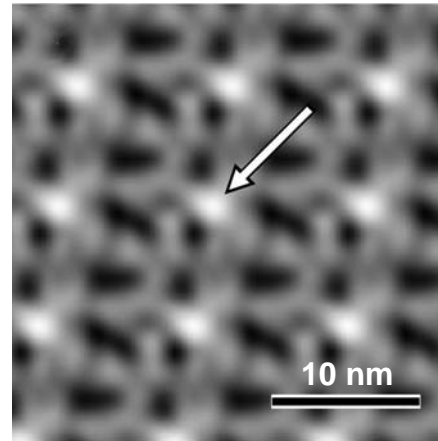
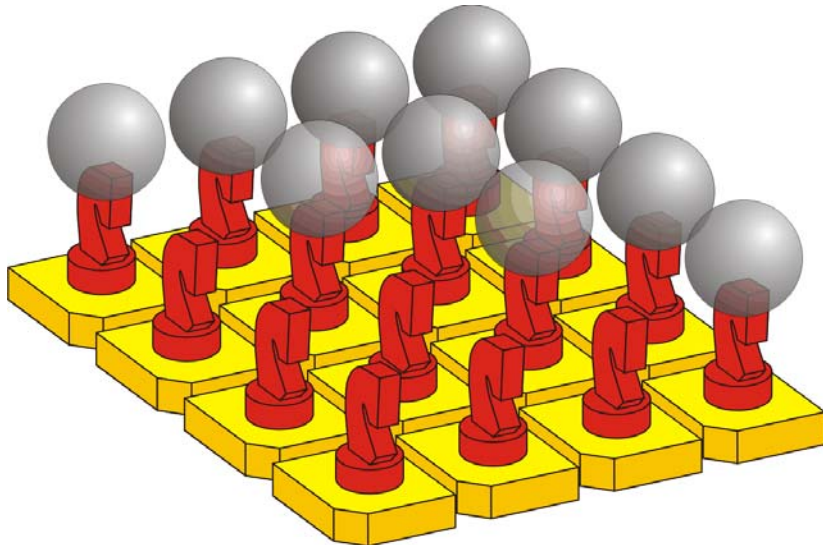
Moll et al., PNAS, 99 (2002) 14646



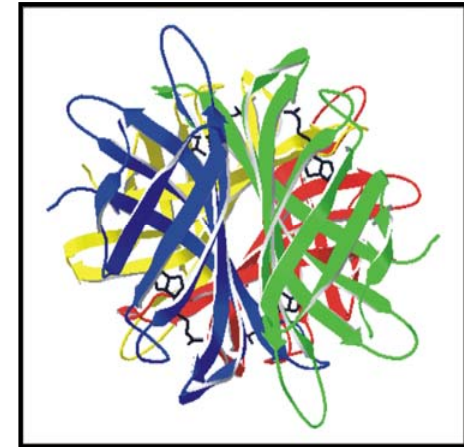
# SbsB / Streptavidin fusion proteins



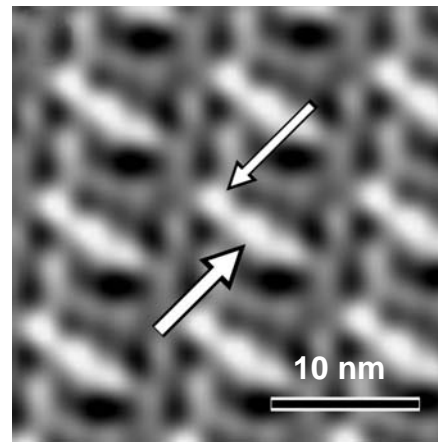
S-layer + function  
(Enzyme, antibody, antigen, ligand)



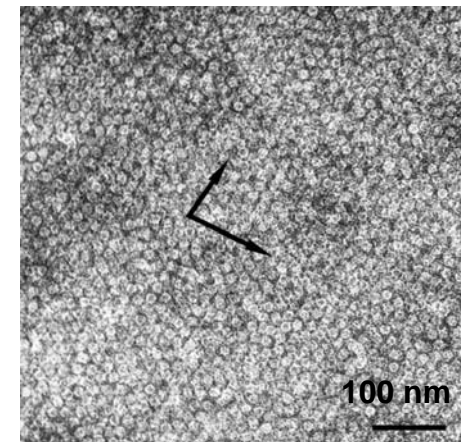
Native S-layer protein



Streptavidin



S-layer fusion protein

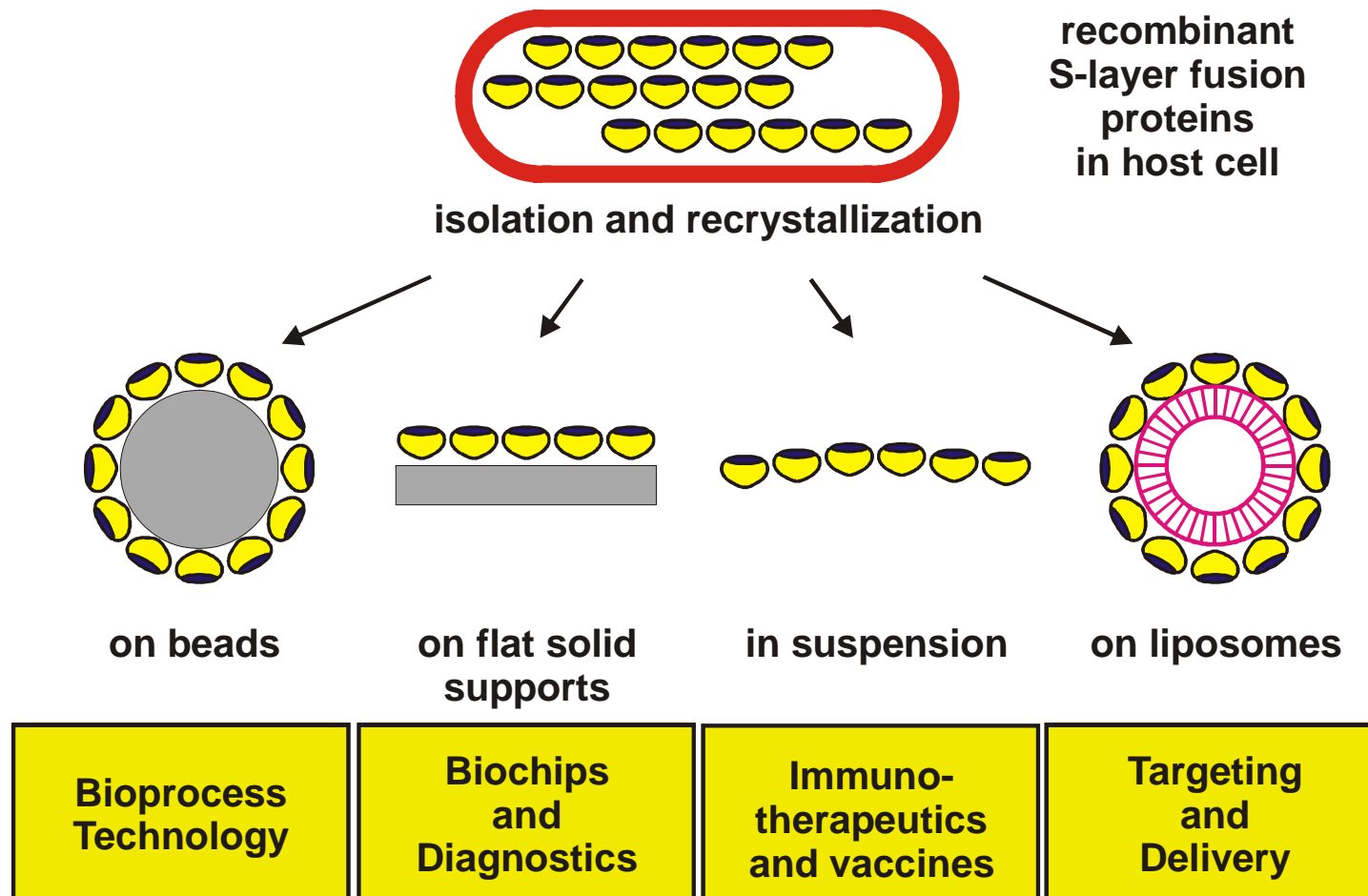


Biotinylated ferritin bound on BS1

# S-layer Fusion Proteins

S-layer fusion protein (selected from various constructs)	Length of funct.	Functionality
rSbsB <sub>1-889</sub> / core streptavidin rSbpA <sub>31-1068</sub> / core streptavidin	118 aa	Biotin binding
rSbpA <sub>31-1068</sub> / Bet v1	116 aa	Major birch pollen allergen
rSbpA <sub>31-1068</sub> / Strep-tag	9 aa	Affinity tag for streptavidin
rSbpA <sub>31-1068</sub> / ZZ	116 aa	IgG-Binding domain
rSbpA <sub>31-1068</sub> / GFP	238 aa	Green fluorescent protein
rSbpA <sub>31-1068</sub> / cAb	117 aa	Heavy chain camel antibody
rSbpA <sub>31-1068</sub> / LamA rSgsE <sub>331-903</sub> / RmlA	263 aa 299 aa	(hyper)thermophilic enzymes
rSbpA <sub>31-1068</sub> / AG4 and AGP35 rSbpA <sub>31-1068</sub> / CO2P2 rSbpA <sub>31-1068</sub> / His rSbpA <sub>31-1068</sub> / rSilC	12 aa 12 aa 6 aa 158 aa	Silver binding Cobalt binding Gold binding Silica & titania binding

# Technologies Based on Recombinant S-layer Fusion Proteins

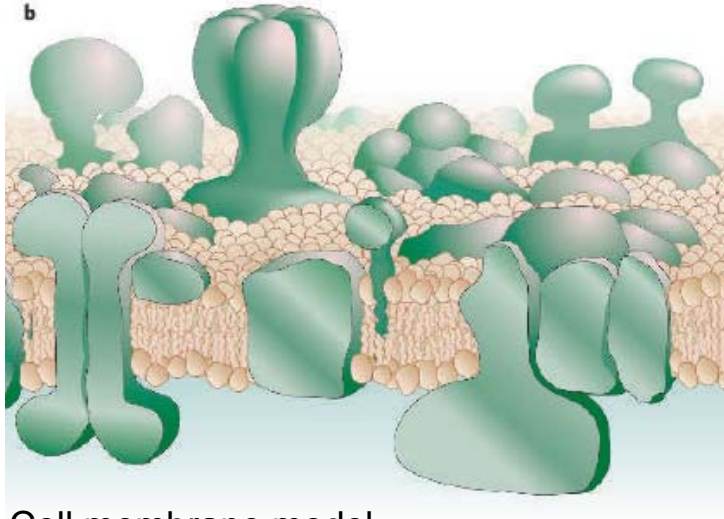




# Applications of S-layer Fusion Proteins

- Sensing layers for label free detection systems (SPR, SAW, QCM-D)
- High density affinity coatings (MDS - blood purification)
- Immunogenic and immunomodulating structures (e.g. development of anti-allergic vaccines)
- Stabilization of Langmuir lipid layers
- Functionalization of liposomes or lipid/plasmid particles as targeting and delivery systems
- Immobilizing of enzymes
- Precipitation and binding of nanoparticles

# Biomimetic Cell Membranes

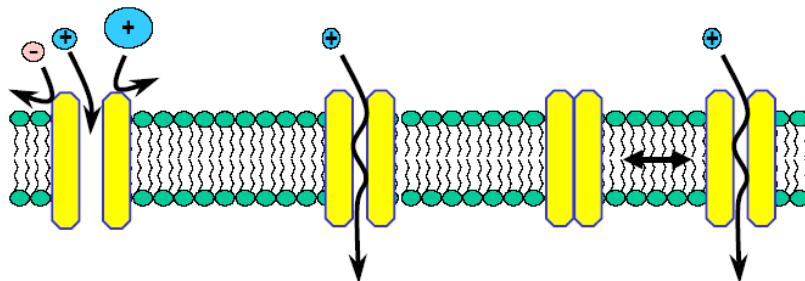


Cell membrane model

Engelman, 2005, *Nature*, 438:578

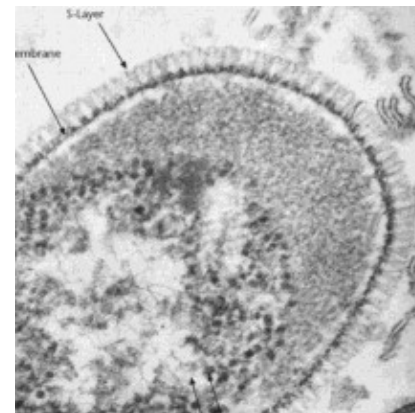
## Specific membrane functions

1.selectivity 2.binding 3.opening / closing

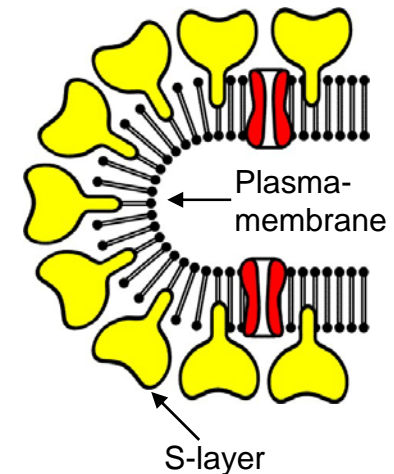


## Archaeal cell envelope structure

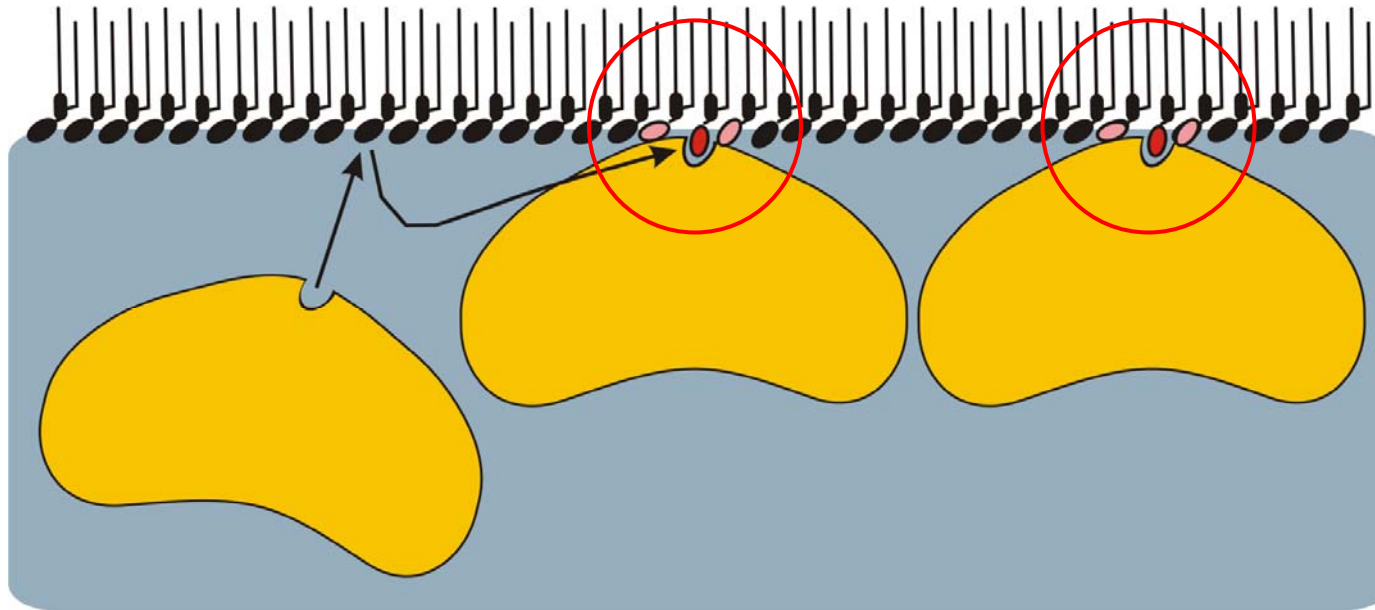
A supramolecular structure optimized in ~ 3,5 billions of years under extreme environmental conditions (120°C, concentrated salt solutions, pH 0, 1100 bar).



TEM of an archaeal cell



# S-layer supported lipid membranes (Semifluid lipid membranes)

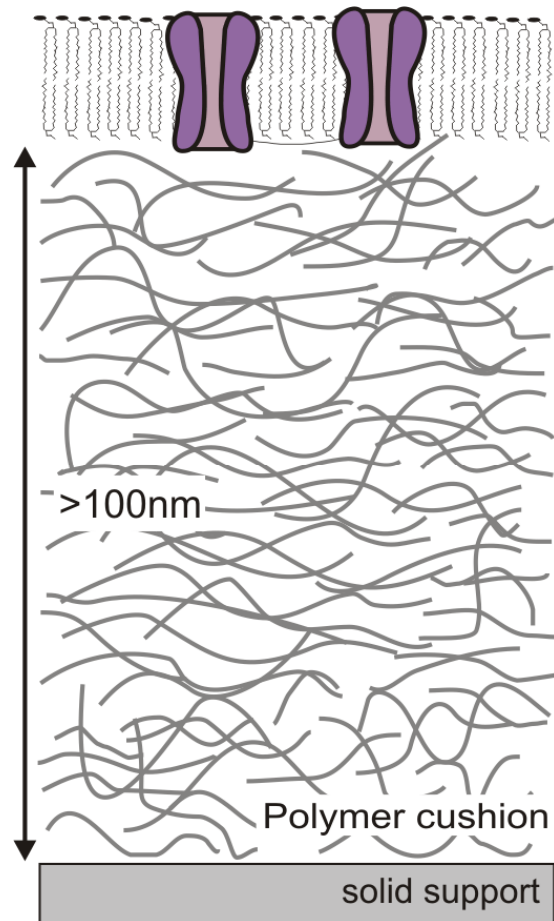


## Methods:

- Grazing-incidence X-ray diffraction
- X-ray and neutron reflectometry
- Infrared spectroscopy (FT-IRRAS)
- FRAP (fluorescence recovery after photo bleaching) with fluorescent lipid probes



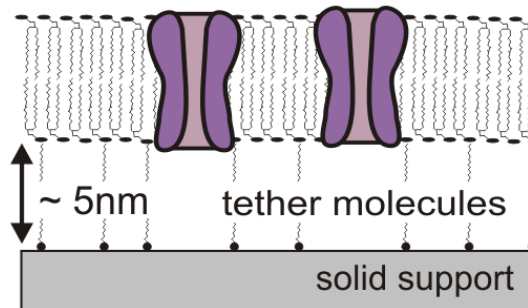
# Biomimetic Lipid Membrane (Semifluid Membrane Model)



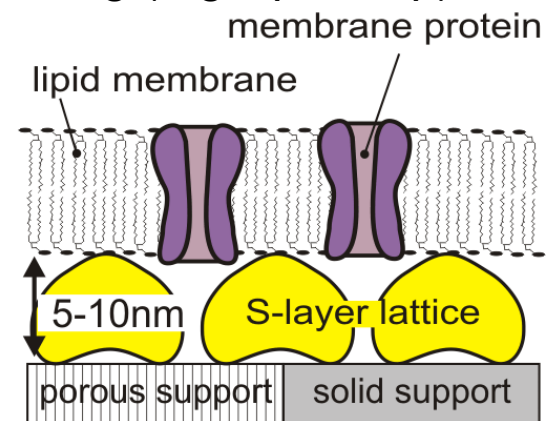
Sackmann, Science 271 (1996) 43

- S-layer stabilized solid supported lipid membranes
- S-layer acts as stabilizing structure, tethering structure and ionic reservoir
- Applications:

Membrane Sensors, Lab-on-a-chip devices,  
High Throughput Screening (e.g. lipid chip),  
DNA sequencing



Schiller et al. Angew. Chem. Int. Ed.  
42 (2003) 208



Schuster et al., Langmuir 17 (2001) 499  
Schuster et al., Langmuir 19 (2003) 2392  
Gufler et al. BBA 1661 (2004) 154

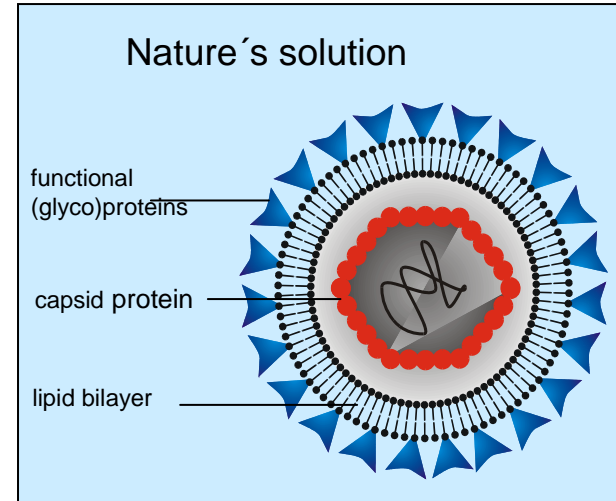
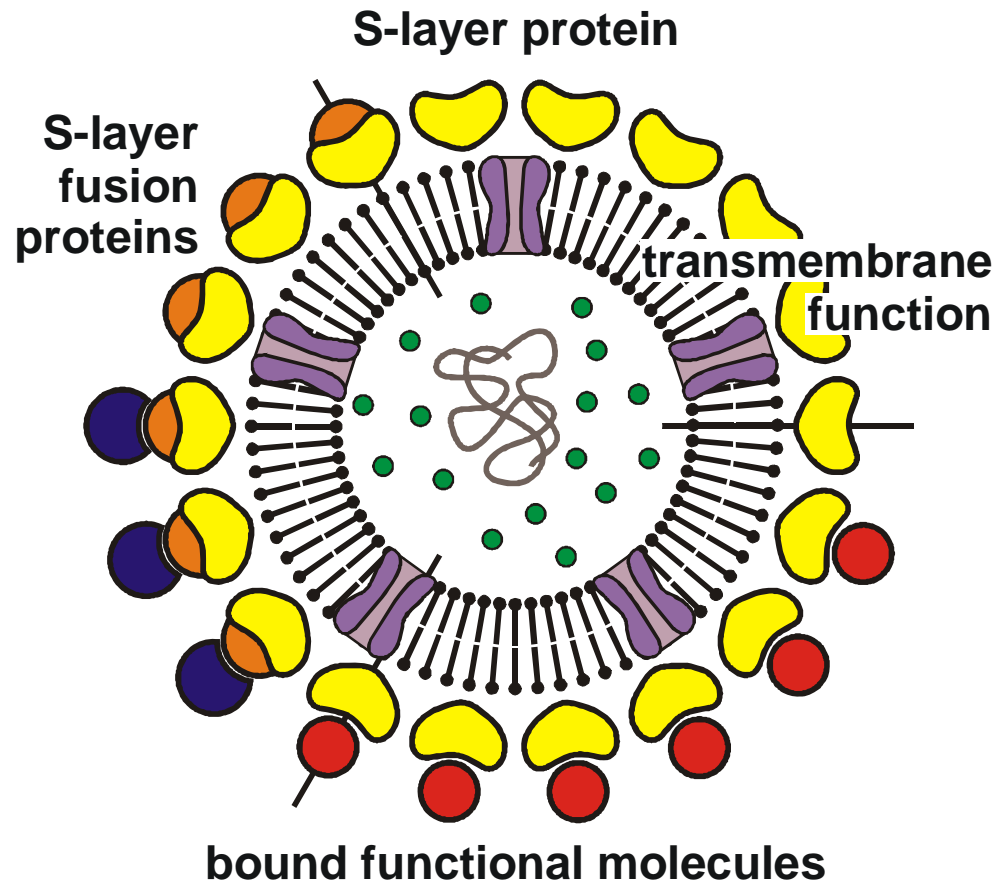
## ...why stabilized Lipid Model Membranes?

- For long-term studies on functional membrane proteins in lipid membranes:
  - ~30 % of all proteins found in various organisms are membrane proteins
  - > 50 % of the proteins interact with membranes
  - ~ 15 % of the most sold drugs act on ion channels
  - ~ 60 % of the ethical drugs affect membrane proteins

### Application potential of functional lipid membranes:

- Diagnostics
- Linking silicon technology and solid state physics with biological systems (e.g. coupling cells to surfaces)
- Lab-on-a-chip, HTScreening
- Biosensors

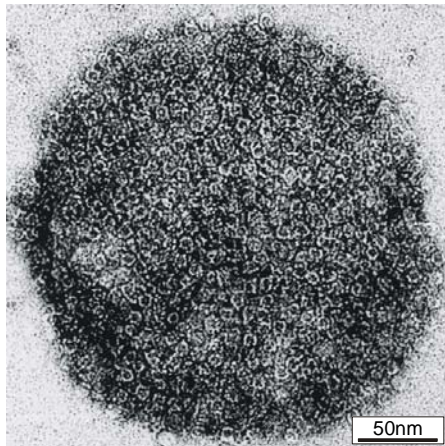
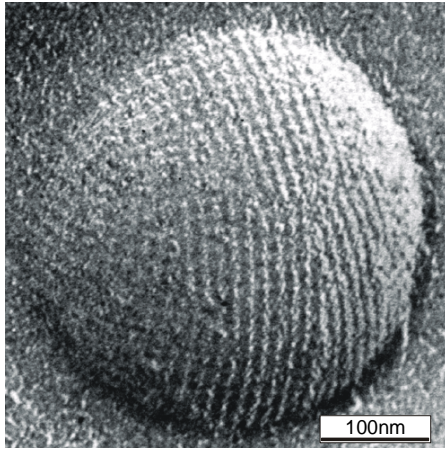
# S-layer coated lipid/plasmid transfection particles (Artificial Virus)



"wooden horse" from the 2004 film *Troy*  
(R. Burgess, GNU Free Doc License)



# Properties of S-layer coated liposomes



What is new in comparison to conventional “naked” liposomes?

- Higher (mechanical and thermal) stability
- Monomolecular coating with functional molecules (e.g. with addressor molecules)
- Possibility for transport of hydrophilic and hydrophobic substances

Potential applications

- Vaccines, immune therapy, drug-targeting and drug-delivery
- Artificial viruses (inclusion of nucleic acid) for gene therapy

Küpcü et al. BBA 1235 (1995) 263  
Mader et al. BBA 1418 (1999) 106  
Mader et al. BBA 1463 (2000) 142

## Summary

Basic research on structure, genetics, chemistry, morphogenesis and function of S-layers has led to a broad spectrum of applications in the Life and Non-life sciences.

The key properties of S-layer proteins for a broad range of applications include their

- ability to self-assemble, and
- to expose functional domains in well defined orientation and position.

# Acknowledgements

Jose-Luis Toca-Herrera	(Biophysical characterization)
Seta Küpcü	(Liposomes)
Dieter Moll	(Streptavidin fusion proteins)
Erika Györvary	(AFM)
Christine Horejs	(MD simulations)
Markus Gossmann	(technician)
Jacqueline Friedmann	(technician)

## **Financial support**

- Austrian Science Fund (FWF)
- Austrian Nano Initiative
- European Union
- Volkswagen Foundation
- Air Force Office of Scientific Research (AFOSR)